



LISTS OF SPECIES

Check List 11(4): 1688, 13 June 2015 doi: http://dx.doi.org/10.15560/11.4.1688 ISSN 1809-127X © 2015 Check List and Authors

Brazilian tropical fishes in their southern limit of distribution: checklist of Santa Catarina's rocky reef ichthyofauna, remarks and new records

Antônio Batista Anderson¹, Alfredo Carvalho-Filho², Renato Araujo Morais¹, Lucas Teixeira Nunes¹, Juan Pablo Quimbayo¹ and Sergio Ricardo Floeter^{1*}

1

- 1 Laboratório de Biogeografia e Macroecologia Marinha, Departamento de Ecologia e Zoologia, Centro de Ciências Biológicas, Universidade Federal de Santa Catarina, Florianopolis, SC 88040-900, Brazil
- 2 Fish Bizz Ltda., Rua Moncorvo Filho, 51, Butantã, São Paulo, SP 05507-060, Brazil
- * Corresponding author. E-mail: sergio.floeter@ufsc.br

Abstract: We present a checklist of 278 species of reef fishes recorded along the coastline of Santa Catarina state, the southernmost limit of distribution of tropical ichthyofauna on the coast of Brazil. Twelve new species records for this state are presented: Acanthurus coeruleus, Acanthurus monroviae, Apogon americanus, Cantherhines macrocerus, Chaetodon sedentarius, Chromis flavicauda, Clepticus brasiliensis, Decapterus punctatus, Gymnothorax vicinus, Herpetoichthys regius, Muraena retifera and Stegastes partitus. Stegastes partitus and H. regius are reported for the first time, respectively, from the Southwestern Atlantic and for the coastal part of this region, while Acanthurus monroviae is reported for the second time for the Southwestern Atlantic. We present habitat distribution, trophic structure and comment on biogeographic affinities of this transitional region, discussing both remarkable species presences and absences.

Key words: rocky reefs, Southwestern Atlantic Shelf, Teleostei, Elasmobranchii, Atlantic Subtropical Convergence, upwelling

INTRODUCTION

The Brazilian reef ichthyofauna has been subject to considerable research in the past 30 years. This was especially due to the popularization of scuba diving among Brazilian scientists, and to the improvements in genetics and computational power (Floeter et al. 2001; Rocha et al. 2008; Bernardi et al. 2013; Pita et al. 2014). However, this region still remains poorly studied in comparison to other biogeographic provinces in the world (Floeter et al. 2001).

The south and southeastern Brazilian coastline is

characterized by granitic rocky reefs influenced by both warm tropical waters from the Brazil Current and cool waters from the South Atlantic Central Water (SACW). This water mass intrudes on the shallow coastal shelf of this region (Acha et al. 2004), especially during spring and summer northeastern winds, and features temperatures of ≤16°C (Carvalho et al. 1998). In the southernmost part of the Brazilian coast, the cold La Plata Plume Water (PPW) coming from the discharge of the La Plata River (at 35°S) reaches coastal areas during the winter (Möller Jr. et al. 2008). The low temperatures generated by these water masses affect the distribution of tropical marine organisms in the region (Boschi 2000; Floeter et al. 2001, 2008; Spalding et al. 2007; Barneche et al. 2009; Anderson et al. 2014a, 2014b), precluding some of them from establishing southwards. Mangrove forests (Sobrinho et al. 1969), corallith (Capel et al. 2012) and rhodolith beds (Gherardi 2004; Pascelli et al. 2013) are biological features of the landscape that reach their southern limit of distribution in the Southwestern Atlantic, precisely in the state of Santa Catarina. Coincidently, this state also represents the southern limit of occurrence of rocky reefs, with a large stretch of sandy beaches extending from it almost continuously to Uruguay. Therefore, for fishes and other organisms that inhabit hard substrates, Santa Catarina is the southernmost limit of the Brazilian biogeographic province (Floeter et al. 2008; Briggs and Bowen 2012).

There are recent taxonomic inventories from São Paulo (Luiz et al. 2008) and Paraná (Hackradt and Félix-Hackradt 2009) states in Brazil, as well as from the coast of Patagonia, in Argentina (Galván et al. 2009). However, despite its biogeographic importance, taxonomic knowledge on Santa Catarina reef fishes remains largely outdated (e.g., Lema 1976; Lema et al. 1980; Godoy

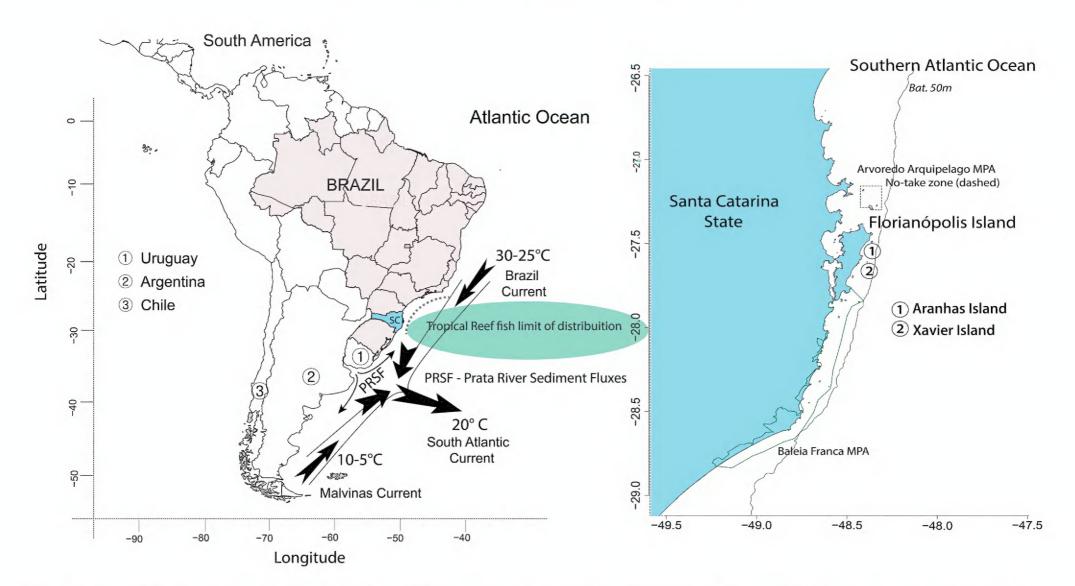


Figure 1. Map of South America showing the influence of both warm tropical and cool waters along the southern Brazilian coast. The Santa Catarina state is represented in light blue (SC). The green ellipse represents the southernmost limit of distribution for tropical reef fish. The dashed arc represents the "Arc of Capricorn" region. The dashed and green polygons and numbers represent most sampled areas.

1987). Only localized (Hostim-Silva et al. 2006) and small-scale initiatives have adressed this issue through the last decade (Barneche et al. 2009; Anderson et al. 2014a). In order to fill this knowledge gap, we provide an updated checklist of reef fishes for the state of Santa Catarina, including twelve new records. Some of these records are largely unexpected given they represent range extensions of many thousands of kilometers over regions where those species were unrecorded, despite the presense of suitable reef habitat.

MATERIALS AND METHODS Study area

The coast of Santa Catarina is located between the latitudes 25°57′ S and 29°23′S, representing approximately 7% of the Brazilian coast (Diehl and Horn Filho 1996) (Figure 1). This region is influenced by continental inputs from rivers in the northern part of the coast (i.e., Itapucu, Itajaí-açu, Tijucas and Tubarão Rivers) (Carvalho et al. 1998; Hille et al. 2008). In the southern portion of the state, the upwelling phenomenon during austral summer and the influence of the La Plata River Plume (see PRSF Figure 1) and Sub-Antarctic Water (Subtropical Shelf Front) during austral winter are key oceanographic processes (Piola et al. 2000; Piola et al. 2005). Complex coastal geography, including numerous coastal islands, and the out put of various small to medium-sized rivers results in various types of environments, each having its own oceanographic features and species assemblages (Charrid 2011).

Data

We base this work on over 12 years of underwater observations using free and scuba diving conducted by the authors (Anderson et al. 2014a; and Marine Macroecology and Biogeography Laboratory photographic data bank), as well as museum vouchers and literature records (i.e., Godoy 1987; Carvalho-Filho 1999; Floeter et al. 2008; Hostim-Silva et al. 2006; Anderson et al. 2014a).

In this paper, we consider reef fish to be those species which are associated with hard substrates after their post-settlement stage, whose habitat includes the continental shelf and islands near the shore, and spend any part of their lifecycle associated with rocky reef systems, including occasional epipelagic and soft substrate species known to occasionally feed, shelter, reproduce or search for cleaning services in rock reefs. Species that have never been observed in reefs in the study region were not considered. This includes species from the families Achiridae, Atherinopsidae, Coryphaenydae, Cynoglossidae, Engraulidae, and some genera of Clupeidae and Scombridae. We consider here species that occur between the surface and depths to 50 m, acknowledging that, albeit deeper occuring species do indeed use reef habitat, we have not been able to adequately sample these depths.

Because of recent changes in the classification of fishes (e.g., Near et al. 2012; Faircloth et al. 2013), fish families are listed alphabetically. We adopted recent taxonomic changes in our classification: Westneat and Alfaro (2005); Craig and Hastings (2007); Smith and Craig (2007); Choat et al. (2012); Boehm et al. (2013); Frable et al. (2013); Knudsen and Clements (2013); and Silveira et al. (2014).

We also included the following information regarding species biology:

Habitat distribution. The physionomy within a rocky reef where a species is usually recorded. We stipulated four different habitat types (Figure 2). The Reef slope (RS) is the zone associated with the presence of rocky substrate ranging from the surface to the point where sediments start to make up a substantial contribution to bottom cover. This reef zone ranges from vertical to gently sloping surfaces and comprise depths varying from six to almost 30 m. The Sandy bottom (SB) is the zone covered essencially by sandy sediments (although silt and clay might also occur in extremely sheltered reefs) adjacent to the rocky reef slope. Carbonate is a minor contributor to these sediments except for a few rhodolith banks that occur in this region. Albeit this could be considered a different zone, fishes that occur in this zone are often the same that occur in sand sediments. Very sparsely scattered granitic boulders also occur in this zone. The Interface (INT) is the transitional zone between the complex rocky reef and the sandy bottom, characterized by hard structures, including some holes, surrounded by a matrix of sand. Water Column (WC) is represented by the pelagic environment adjacent to the rocky reef (adapted from Luiz et al. 2008).

Abundance indicator. Based on a diver's likelihood of recording a species in its usual habitat and depth range on any given dive (adapted from Feitoza et al. 2003; Luiz et al. 2008; Humann and DeLoach 2014), where CO = common (sightings are frequent);

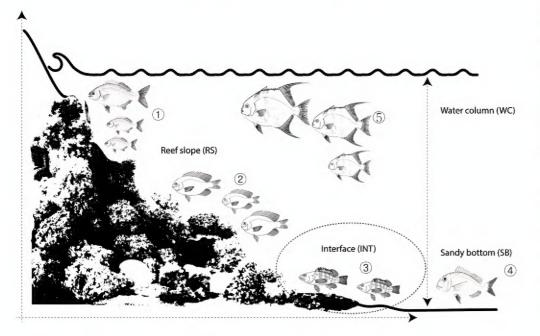


Figure 2. Hypothetical Santa Catarina rocky reef with examples of reef fish species typically associated with different zones. The Reef slope (RS) is associated to hard substrate, the Sandy bottom (SB) to sediments, the Interface (INT) is a transitional zone between the RS and the SB; and the Water column (WC) is absent on substrate. Examples of species commonly associated with a specific zone: (1) *Kyphosus vaigiensis* and (2) *Stegastes fuscus* (RS); (3) *Serranus flaviventris* (INT); (4) *Calamus penna* (SB) and (5) *Chaetodipterus faber* (WC).

OC = occasional (sightings are not expected on a regular basis); UN = unusual (sightings occur less than occasionally); and RA = rare (sightings are exceptional).

Geographic range. The ranges of occurrence for species were based primarily in Floeter et al. (2008) and Carvalho-Filho (1999), with additional notes provided by Galván et al. (2009). Abbreviations are as follow CT = Circumtropical; CG = Circumglobal; AO = Atlantic Ocean; TA = Tropical Atlantic; EA = Eastern Atlantic; WA = Western Atlantic; SWA = Southwestern Atlantic; NWA = Northwestern Atlantic; MAR = Mid-Atlantic Ridge; MED = Mediterranean Sea; WIO = Western Indian Ocean; IP = Indo-Pacific Ocean; NWP = Northwestern Pacific; and TEP = Tropical Eastern Pacific (Froese and Pauly 2014). Brazilian Province endemics include species recorded from the southern tip of the Caribbean (Venezuela, Trinidad and Tobago and other islands of the Lesser Antilles), Cape Verde Archipelago and Ascension Island (Freitas et al. 2014), but which have 90% or more of its range in Brazil.

Trophic category. The diet of a species was based both in the literature (Randall 1967, 1996; Carvalho-Filho 1999; Ferreira et al. 2004; Luiz et al. 2008) and indirect observations performed by the authors, where MCAR = Macrocarnivores (species which feed mainly on mobile organisms, such as macroinvertebrates and fishes); MINV = Mobile Invertebrate Feeders (species which feed primarily on benthic mobile invertebrates, such as mollusks, crustaceans, and worms associated with hard or nearby unconsolidated substrate); OMNI = Omnivores (species which feed on a variety of resources, but that necessary include invertebrates and algae); PLANK = Planktivores (species which feed primarily on macro- and microplankton); HERB = Herbivores/ Detritivores (species, both nonterritorial and territorial herbivores, which include in their diet detritus and macroalgae) and SINV = Sessile Invertebrate Feeders (species which feed on sessile benthic invertebrates, such as cnidarians, bryozoans, ascidians and sponges).

Record type. The method by which species were recorded and documented: VOU = Museum Vouchers (the institutions and voucher numbers of specimens are provided in Appendix 1); LIT = Literature; PHO = Photographs and SIG = Sighting during underwater fieldwork.

Multivariate analysis. To describe associations of fish families to trophic categories and habitat distribution we employed a Correspondence Analysis (Nenadic and Greenacre 2007) based on species richness (i.e., number of species per family). To avoid distortions caused by highly over-dispersed data, a "Hellinger" transformation was applied before proceeding with statistical analysis (Greenacre 2007).

RESULTS AND DISCUSSION

A total of 278 reef fish species in 170 genera and 74 families have been recorded along the coast of Santa Catarina during the past 12 years of underwater observations, as well as from the literature and museum vouchers (Table 1, Figure 3).

Based on species richness, the most representative families were Carangidae (20 species), Labridae (19 species), Carcharhinidae (11 species) and Epinephelidae (10 species). The most species-rich genera were *Carcharhinus* (eight species), followed by *Sphoeroides* and *Sparisoma* (five species). A total of 73 species were considered as "common" (CO = 26.3%), 69 species were considered "occasional" (CO = 24.8%), 132 species were considered "rare" (CO = 24.8%), 132 species were considered "rare" (CO = 24.8%) (Figure 4), and four species were considered "unusual" (CO = 24.8%).

Some species are considered as "resident" organisms in the rocky reef systems, which means that they are dependent on the rocky reefs to complete their life cycles (62.6% or 174 species). All the others spend only part of their lives inhabiting the rocky reefs or adjacent habitats, and are able to survive using other habitats.

Trophic Structure

The two dominant trophic groups in this coastal region were the mobile invertebrate feeders (38.8%) and the macrocarnivores (32.4%), followed by planktivores (8.6%), omnivores (8.3%), herbivores/detritivores (7.6%) and sessile invertebrate feeders (4.3%) (Figure 5).

The high proportion of the mobile invertebrate feeders is a characteristic of reef fish assemblages worldwide (Ferreira et al. 2004; Luiz et al. 2008). The predators herein referred to as macrocarnivores include mainly Carcharhinidae (sharks), Carangidae (jacks and pompanos), Epinephelidae (groupers), Lutjanidae (snappers) and Scombridae (tunas and mackerels). In Santa Catarina, most planktivore species are Clupeidae and of the genus Chromis (Pomacentridae), as well as few species from other families. The herbivore/detritivores are mainly represented by Pomacentridae and Labridae-Scarini species. The sessile invertebrate feeders in this rocky environment are the generalists Chaetodontidae and Pomacanthidae, which consume a considerable amount of cnidarians and sponges, respectively. These families are also known to rely heavily on mobile invertebrates and algae, respectively.

Habitat distribution and threatened species

Distributions of rocky reef species within the habitat types, as evidenced herein by the Correspondence Analysis, mirror the classic ecological partitioning by fishes of the Brazilian rocky reef habitats (Sazima 1986). Specifically, apex predators, such as sharks, mesocarnivores, such as Carangidae and Scombridae, as well as Clupeidae and Engraulidae planktivores,

all occupy the water column strata. Herbivores/detritivores, omnivores, mobile invertebrate feeders and Epinephelidae mesocarnivores occupy the reef slope, while mullets and flat fishes dwell on the sandy bottom (Sazima 1986, Figure 6).

Several species are considered threatened according to the IUCN endangered species Red List (IUCN 2015). The relative proportion of threatened species has reached 8.3% (or 23 of 278 species) and encompasses mostly top predators, such as sharks (34.8%) and groupers (17.4%).

Taxonomic updates

Following recent revisions of the families Kyphosidae (Knudsen and Clements 2013), Scaridae (Westneat and Alfaro 2005; Choat et al. 2012) and Serranidae (Craig and Hastings 2007; Smith and Craig 2007), and the genera Hippocampus (Boehm et al. 2013) and Synodus (Frable et al. 2013), some taxonomic updates shall be discussed. The whole family Scaridae is now recognized as a lineage of Labridae and now represents the Tribe Scarini (Westneat and Alfaro 2005; Choat et al. 2012). The family Epinephelidae was split from Serranidae, and the snowy grouper complex Epinephelus niveatus is now included in the previously invalidated genus Hyporthodus (i.e., Hyporthodus niveatus).

In the family Kyphosidae, the former species Kyphosus incisor (Cuvier, 1831), listed from Santa Catarina along with *Kyphosus sectatrix* (Linnaeus, 1758), both by Carvalho-Filho (1999) and Hostim-Silvaet al. (2006), is now revalidated as Kyphosus vaigiensis (Quoy & Gaimard, 1825) (Knudsen and Clements 2013). As this revision is recent, we could not determine wether the other Atlantic species, K. cinerascens and K. bigibbus do occur in Santa Catarina. In the genus Hippocampus, the Southwestern Atlantic species of the *H. erectus* complex is now considered to harbor two species, being *H. erectus* restricted to Brazil (Silveira et al. 2014), and the other ranging from Brazil to Uruguay and Argentina under the name of Hippocampus patagonicus Piacentino & Luzzatto, 2004 (Boehm et al. 2013; Silveira et al. 2014). In the Synodus genus, Synodus foetens (Linnaeus, 1766) may be considered restricted to nortwestern Atlantic (Frable et al. 2013). The species recorded for the southern part of Caribbean is now renamed as Synodus bondi Fowler, 1939. Although the authors suggest that the species which occur along Brazilian coast should be S. bondi (and we therefore consider it to), they did not possess a sufficient sampling of Brazilian individuals to avert the possibility that it is yet another species.

New Records and extreme range extensions

During this work, twelve species of reef fish were recorded for the first time for the coast of Santa Catarina: *Acanthurus coeruleus* (Bloch & Schneider, 1801); *Acanthurus monroviae* Steindachner, 1876; *Apogon*

Table 1. Checklist of reef fish species recorded at Santa Catarina State, Southern Brazil. The genera and species are separate in Chondrichthyes and Actinopterygii arranged in alphabetical order within families. IUCN Status: CR = Critically Endangered (extremely high risk of extinction in the wild); EN = Endangered (high risk of extinction in the wild); NT = Near Threatened = Least Concern (Lowest risk); **DD** = Data Deficient (not enough data to make an assessment of its risk of extinction); **NE** = Not Evaluated (not yet evaluated). **Irophic Category: MCAR** = Carnivore; **MINV** = Mobile Invertebrate Feeder; **SINV** = Sessile Invertebrate Feeder; **OMNI** = Omnivore; **PLANK** = Planktivore; **HERB** = Herbivore/Detritivore. **Habitat: RS** = Reef Slope; **INT** Residence (RE): R = Reef associated. Occurrence: CO = Common; OC = Occasional; UN = Unusual; RA = Rare. Geographic range: CT = Circumtropical; CG = Circumglobal; AO = Atlantic Ocean; TA = Tropical Atlantic; EA = Eastern Atlantic; WA = Western Atlantic; SWA = Southwestern Atlantic; NWA = Northwestern Atlantic; MAR = Mid-Atlantic Ridge; MED = Mediterranean Sea; WIO = Western Indian Ocean; IP = Indo-Pacific Ocean; NWP = Northwestern Pacific; TEP = Tropical Eastern Pacific. Record Type: LIT = in litteris; VOU = Museum Voucher; PHO = Photograph; SIG = Underwater sighting. * = New record; + = Brazilian endemic species, including species recorded in southern tip of the Caribbean, Cape Verde Archipelago and Ascension Island (see Freitas et al. 2014). (likely to become endangered in the near future); LC = Interface; **SB** = Sandy Bottom; **WC** = Water Column.

Circle chirchindia Concentioned controls Mobile 8 Febreis 1839) IV MCAR WC NA MCARAMEDIP VOD Conclusionate decoration Mobile 8 Febreis 1839) IV MCAR WC RA CT IV Conclusionate decoration Mobile 8 Febreis 1839) IV MCAR WC RA CT IV Conclusionate decoration Mobile 8 Febreis 1839) IV MCAR WC RA CT IV Conclusionate decoration Graduations optimized a popula Graduations optimized a popula WC RA CMA VOD Conclusionate decoration Graduationates approared a febreis 1839) IV MCAR WC RA WA VOD Conclusionates optimized Graduationates approared a febreis 1839) IV MCAR WC RA WA VOD Displaying treatment Graduationates approared a febreis 1839) IV MCAR WC RA WA VOD Displaying treatment Graduationates approared a febreis 1839) IV MCAR	Family	Species	Authority	IUCN	Trophic	Habitat	REO	Occur.	Geog. range	Rec.Type
Cardiophylatic stooden Midlier 8 Heale, 1839 LT MiCAR WC CD Cardiophylatic stooden (Mullier 8 Heale, 1839) NT MiCAR WC RA CT Cardiophylatic stooden (Mullier 8 Heale, 1839) NT MiCAR WC RA CT Cardiophylatic stooden (Mullier 8 Heale, 1839) VU MiCAR WC RA CT Cardiophylatic splittler (Mullier 8 Heale, 1839) VU MiCAR WC RA WA Cardiophylatic splittler (Peay, 1832) D MiCAR WC RA WA Cardiophylatic splittler (Peay, 1832) D MiCAR WC RA WA Dayyotic survicious (Midler 8 Heale, 1832) D MiCAR WC RA WA Dayyotic survicious (Midler 8 Heale, 1832) L MiCAR WC RA WA Dayyotic survicious (Midler 8 Heale, 1832) L MiCAR WC RA WA Dayyotic survicious (Midler 8 S	Carcharhinidae	Carcharhinus brevipinna	(Müller & Henle, 1839)	Z	MCAR	WC	8	A	WA/EA/MED/IP	VOU
Cardiorhinous binducties Muller & Heinle 1839) NT MACAR WC RA CT Cardiorhinous binducties Cardiorhinous binducties (Lebuer, 1818) NT MCAR WC RA WAFVANED/IP Cardiorhinous binducties (Lebuer, 1818) NT MCAR WC RA WAFVANED/IP Cardiorhinous potenties (Pearly 1827) VD MCAR WC RA WAFVANED/IP Cardiorhinous potenties (Pearly 1827) VD MCAR WC RA WAFVANED/IP Cardiorhinous potenties (Pearly 1827) NT MCAR WC RA WAFVANED/IP Cardiorhinous potenties (Pearly 1825) NT MCAR WC RA WAFVANED/IP Cardiorhinous potenties (Pearly 1825) NT MCAR WC RA WAFVANAD Cardiorhinous potenties (Pearly 1825) NT MCAR WC RA WAFVANAD Dayote potenties (Pearly 1821) NT MCAR WC RA WAFVANAD		Carcharhinus isodon	(Müller & Henle, 1839)	C	MCAR	WC	0	Q	WA	NOU
Conclusional systems Conclusional systems NT MCAR WC RA CT Conclusional systems (Lassieut, 1818) VU MCAR WC RA VMCAR Conclusional systems (Narch, 1827) VU MCAR WC RA CT Conclusional systems (Renard, 1837) DD MCAR WC RA VMCAR Conclusional systems (Renard, 1839) DD MCAR WC RA VMCAR Gordecendo cunier (Renard, 1839) DD MCAR WC RA VMCAR Gordecendo cunier (Renard, 1839) DD MCAR WC RA VMCAR Dosyatis consentant (Moller & Berlin, 1839) DD MCAR WC RA VMCAR Dosyatis contractor (Moller & Berlin, 1839) DD MCAR WC RA VMCAR Dosyatis contractor (Moller & Berlin, 1839) DD MINV SB RA VMA Dosyatis propositis propositis contractor (Moller		Carcharhinus leucas	(Muller & Henle, 1839)	Z	MCAR	WC	~	A	Ь	П
Cardenthinise because Cardenthinise because Cardenthinise because (Value or 1827) VU MCAR WC RA WWEANDIP Cardenthinise pointies (Natide) 1827) VU MCAR WC RA WA Cardenthinise pointies (Percia 1828) DV MCAR WC RA WA Cardenthinise pointies (Percia 1828) DV MCAR WC RA WA Rhizophonocden jonnels (Percia 1839) DV MCAR WC RA WA Rhizophonocden jonnels (Percia 1851) LC MCAR WC RA WA Dosystic specialization (Percia 1859) LC MCAR WC RA WA Dosystic specialization (Percia 1852) LC MINV SB RA WA Dosystic specialization (Robal 24) C MINV SB RA WA Dosystic specialization (Robal 24) LC MINV SB RA WA Dosystic specializat		Carcharhinus limbatus	(Muller & Henle, 1839)	LN L	MCAR	WC	~	A	Ь	ПТ
Cardedninus glunteus (Nardo 1827) VU MCAR WC RA WA Cardenthinus glounes (Ranzani 1839) DD MCAR WC RA WA Cardenthinus glounes (Poey, 1868) V WC RA WA Cardenthinus signate (Poey, 1868) V WC RA WA Galeceredo cuvier (Poey, 1861) DD MCAR WC RA WA Alteracionacion ponosus (Poey, 1861) DD MINA SB RA WA Alteracionacion ponosus (Poey, 1861) DD MINA SB RA WA Dosyatis centroura Hildebrand & Schroeder, 1928 DD MINA SB RA WA Dosyatis centroura Hildebrand & Schroeder, 1929 LC MINA SB RA WA Dosyatis durinacia (Bioch & Schroeder, 1923) LC MINA SB RA WA Agratia durinacia (Bioch & Schroeder, 1923) LC MINA SB		Carcharhinus obscurus	(LeSueur, 1818)	N	MCAR	WC	~	A	WA/EA/MED/IP	VOU
Carcharbinus poresus (Hanchari, 1839) DD MCAR WC RA WMEA Carcharbinus poresus (Peore, 1828) 1, 0 MCAR WC RA WMEA Galescerbo curvier (Peore, 1828) 1, 0 MCAR WC RA WMEA Ribzoprionodan parasa (Peore, 1831) 1, 0 MCAR WC RA WA Ribzoprionodan parasa (Peore, 1861) 1, 0 MCAR WC RA WA Dosystis controura (Mileche Heine, 1831) 1, 0 MINV SB R RA WA Dosystis controura (Mileche Schneder, 1801) DD MINV SB R R WA Dosystis controura (Mileche Schneder, 1801) DD MINV SB R R WA Dosystis controura (Bonoparte, 1832) DD MINV SB R R WA Dosystis propertice (Bonoparte, 1831) VU MINV SB R R C <t< td=""><td></td><td>Carcharhinus plumbeus</td><td>(Nardo, 1827)</td><td>N</td><td>MCAR</td><td>WC</td><td>~</td><td>А</td><td>Ь</td><td>LIT/VOU</td></t<>		Carcharhinus plumbeus	(Nardo, 1827)	N	MCAR	WC	~	А	Ь	LIT/VOU
Carcharbinus signatus (Poep, 1868) VI MICAR WC RA WM-EA Galescende condever (Poep, 1861) 187 MICAR WC RA WM-EA Rhizoparionaden Jaconsius (Poep, 1861) LC MICAR WC RA WM Albizoparionaden Josensius (Poep, 1861) LC MICAR WC RA WM Dasystis annicaria (Hildier & Heinel, 1832) LC MINV SB R RA WA Dasystis cantoura (Bloch & Schneder, 1801) LD MINV SB RA WA Dasystis cantoura (Bloch & Schneder, 1801) LC MINV SB RA WA Dasystis cantoura (Bloch & Schneder, 1801) LC MINV SB RA WA Dasystis cantoural clinede (LILINAGER, 1802) LC MINV SB RA WA Sentral Reposition (LILINAGER, 1803) LC MINV SB RA WA Sentral Reposition (LILI		Carcharhinus porosus	(Ranzani, 1839)	QQ	MCAR	WC	~	A	WA	NOU
Gelecoration tower (Peroin &LeSueur,1822) NT MCAR WC RA CT Phizzapionodou Indundii (Waller & Henie, 1839) LC MACAR WC RA WA Phizzapionodou Indundii (Yeay, 1861) LC MINV SB R RA WA Posydis cantoura (Mitchil, 1815) LC MINV SB R RA WA Dosydis cantoura (Mitchil, 1815) LC MINV SB R RA WA Dosydis cantoura (Mitchil, 1815) LC MINV SB R RA WA Dosydis cantoura (Bich & Schneder, 1810) DD MINV SB R RA WA Dosydis cantoura (Bich & Schneder, 1810) VU MINV SB R RA CG Antonia (Bich & Schneder, 1810) VU MINV SB R R RA CG Antonia (Bich & Schneder, 1810) VU MINV SB R		Carcharhinus signatus	(Poey, 1868)	N	MCAR	WC	~	A	WA/EA	VOU
Physicaphionodan lalandii (Muller & Henle, 1839) DD MCAR WC RA WA Panyagatis americana (Peey, 1811) 1 C MCAR WC RA WA Dosyadis centroura Hildebanda & Schroeder, 192B D MINV 5B RA WA Dosyadis centroura (Mirthill, 1815) LC MINV 5B RA WA Dosyadis centroura (Hildebanda & Schneder, 1801) LC MINV 5B RA WA Dosyadis centroura (Bloch & Schneder, 1801) LC MINV 5B RA WA Persopatis processor (Bloch & Schneder, 1812) LC MINV SB RA WA Astrono control (Bloch & Schneder, 1812) LC MINV SB RA WA Astrono control (Bloch & Schneder, 1812) LC MINV SB RA WA Astrono control (Bloch & Schneder, 1812) NT PLANK WC LN CG Astrono control (Gulfre, 1		Galeocerdo cuvier	(Perón &LeSueur, 1822)	Ŋ	MCAR	WC	~	А	Ь	П
Philogophionodop porosus (Poey, 1861) LC MCAR WC RA WAA Dosyatis enrictona Hildeband & Schoeder, 1928 DD MINV SB R RA WAA Dosyatis enrictona (Mitchil, 1813) DD MINV SB R RA WAS Dosyatis enrictona (Bloch & Scheeder, 1801) DD MINV SB RA WAS Dosyatis guttata (Bloch & Scheeder, 1801) LC MINV SB RA WAS Percoplostyrigon violicea (Bloch & Scheeder, 1810) VJ MINV SB RA WAS Simus opythistics (Euphrasen, 1790) NT MINV SB RA GG Aerobatus natinati (Euphrasen, 1790) NT MINV WC RA GG Aerobatus natinati (Euphrasen, 1790) NT PLANK WC RA GG Aerobatus natinati (Euphrasen, 1790) NT PLANK WC RA GG Aerobatus <		Rhizoprionodon lalandii	(Müller & Henle, 1839)	DD	MCAR	WC	~	А	WA	VOU
Dasyactis americana Hildebrand & Schroeder, 1928 DD MINV SB RA WAFA Dasyaris centroura (Mirchill, 1815) LC MINV SB RA WAFA Dasyaris centroura (Block & Schneider, 1801) DD MINV SB RA WAFA Dasyaris centroura (Block & Schneider, 1801) LC MINV SB RA SWA Percoplaty rogan violecze (Block & Schneider, 1832) LC MINV SB RA CG Sumus oxylinicus (Brineseus, 1788) VU MCAR WC RA CG Aerdechus ranicari (University) VU MCAR WC RA CG Adechus trustoni (Lloyd, 1908) NT PLANK WC NC RA Mounter brasilensis (Olers, Ball) DD MINV SB R C Acadabus trustoni (Universide, 1831) DD MINV SB R C Acadamicos porteuirs (Olers, Ball) (Ole		Rhizoprionodon porosus	(Poey, 1861)	C	MCAR	WC	~	A	WA	ПТ
Dasyastiz centroura (Mitchill, 1815) LC MINV SB RA WA/EA Dasyastiz guttata (Borb & Schneider, 1801) DD MINV SB RA CMA Dasyastis pytostiqua (Boneparte, 1812) LC MINV SB RA CG Perceptaty/yegon violocea (Unnaeus, 1758) VU MINV SB RA CG Isuus coyribricus Refinesque, 1810 VU MINV SB RA CG Areabetus soninari (Lubriassen, 1790) NT PLANK WC RA CG Mobula thurstoni (Libria, 1908) NT PLANK WC NN CG Anothie sturus Refinesque, 1810 VU MARN SB RA CG Rehinobaros percelleras (Olfers, 1831) DD MINV SB RA CG Rehinobaros percelleras (Vallabaun, 1792) NT MINV SB RA CG Sphyma levinis (Uliria, Berlei, 1841) VU	Dasyatidae	Dasyatis americana	Hildebrand & Schroeder, 1928	QQ	MINV	SB		A	WA	ПТ
Dasyatis guttata (Bloch & Schneider, 1801) DD MINV SB RA WA Dasyatis pytostigma Santos & Carvalho, 2004 LC MINV SB RA SWA Petroplatyrygan violacea (Bonaparte, 1832) LC MINV SB RA CG Gymmura altrated (Linnaeus, 1758) VU MCAR WC RA CG Aetobatus narinari (Euphrasen, 1790) NT MINV WC RA CG Mobula thustsoni (Luoyd, 1908) NT PLAMK WC R RA CG Mobula thustsoni (Luoyd, 1908) NT PLAMK WC N CG Mobula thustsoni (Luoyd, 1908) NT PLAMK WC N CG Acarcharios teurus Rahinobatos percellens (Muller & Henle, 1841) VU MCR RA CG Rhinobatos percellens (Muller & Henle, 1841) VU MCAR WC RA CG Sphyma telunin (Griffith & Smith, 1		Dasyatis centroura	(Mitchill, 1815)	C	MINV	SB	~	А	WA/EA	SIG/LIT/PHO ³
Dasyatis hypostigma Santos & Carvalho, 2004 LC MINV SB RA SWA Piercoplatyrygan violacea (Bonaparte, 1832) LC MINV SB R RA CG Sumua actualed (Linnaeus, 1758) VU MINV SB R RA CG Aerobaus natinati (Euphrasen, 1790) NT MINV WC R RA CT Mobula thurstori (Lino(1908) NT PLANK WC R RA CG Mobula thurstori (Lio/(1908) NT PLANK WC R R CG Acriche basilersis (Ulo/(1908) NT PLANK WC R CG Acriche basilersis (Ulo/(1908) NT PLANK WC R R CG Rhinobatos percellers (Waller & Henle, 1841) VU MAAR WC RA CG Sphyma elwin (Gilfith & Smith, 1834) EN MAAR WC RA CG <td< td=""><td></td><td>Dasyatis guttata</td><td>(Bloch & Schneider, 1801)</td><td>DD</td><td>MINV</td><td>SB</td><td>8</td><td>А</td><td>WA</td><td>SIG/LIT/PHO^{1,3}</td></td<>		Dasyatis guttata	(Bloch & Schneider, 1801)	DD	MINV	SB	8	А	WA	SIG/LIT/PHO ^{1,3}
Pteroplacytrygon violacea (Bonaparte, 1832) LC MINV SB RA CG Symmura altavela (Linnaeus, 1758) VJ MCAR WC RA CG Aetodatus aninari (Euphrasen, 1790) NT MINV WC NA CG Marta bursatis (Walbaru, 1792) NT PLANK WC UN CG Marta bursatis (Usod, 1908) NT PLANK WC UN CG Marta bursatis (Usod, 1908) NT PLANK WC UN CG Marta bursatis (Olfers, 1831) DD MINV SB R CG Rhinobaros horkellis Muller & Henle, 1841 VJ MCAR WC NA SG Rhinobaros horkellis Muller & Henle, 1841 VJ MINV SB RA SA Rhinobaros horkellis Muller & Henle, 1841 VJ MINV SB RA SWA Sphyma evinis Griffith & Smith, 1828 VJ MCAR		Dasyatis hypostigma	Santos & Carvalho, 2004	C	MINV	SB	~	A	SWA	VOU
Gymmura altavela (Linnaeus, 1758) VU MINV SB RA WAZEA Surus oxythincus Rafinesque, 1810 VU MCAR WC RA CG Aerobatus narinari (Euphrasen, 1790) NT MINV WC N CG Monta birostris (Walbaum, 1792) NT PLANK WC N CG Monta birostris (Olfers, 1831) DD PLANK WC N CG Advicine burius (Olfers, 1831) DD PLANK WC/RS R CG Advincine burius (Olfers, 1831) VU MCAR WC/RS R CG Rhinobactos parcellens (Walbaum, 1792) NT MINV SB R RA SWA Rhinodatos percellens (Walbaum, 1792) NT MINV SB R RA CG Sphyrand töburo (Unimeeu, 1841) VU MCAR WC RA CG Sphyrand töburo (Linnaeus, 1758) LC		Pteroplatytrygon violacea	(Bonaparte, 1832)	PC	MINV	SB	R	А	CG	SIG/LIT/PHO ³
Aetobauts nativation Rafinesque, 1810 VU MCAR WC RA CG Aetobauts nativati (Euphrasen, 1790) NT MINV WC R AC Mobula thuistorii (Ulodi, 1908) NT PLANK WC UN CG Mobula thuistorii (Ulodi, 1908) NT PLANK WC NN CG Marcine biosstiis (Olfers, 1831) DD MINV SB R CG WA Antribobatos brokelii Muller & Henle, 1841 CR MINV SB RA CG Ashinobatos percellens (Walbaum, 1792) NT MINV SB R RA CG Aphyma leviniotsris (Muller & Henle, 1841) VU MINV SB R RA CT Sphyma leviniotsris (Muller & Smith, 1828 VU MCAR WC RA CG Sphyma levinio (Ilimaeus, 1758) LC MCAR WC RA CG Sphyma levinio (Ilimaeus, 1	Gymnuridae	Gymnura altavela	(Linnaeus, 1758)	۸n	MINV	SB		А	WA/EA	SIG/LIT/PHO¹
Aetobatus narinari (Euphrasen, 1790) NT MINV WC R RA CT Manta birostris (Walbaum, 1792) NT PLANK WC UN CG Mobula thurstorii (Lloyd, 1908) NT PLANK WC UN CG Narche brasiliensis (Olfers, 1831) DD MINV SB R OC WA Rhinobaros rorkelir Muller & Henle, 1841) VJ MCAR SB RA SWA Rhinobaros percellens (Walbaum, 1792) NT MINV SB R AC WA Rhinodaros percellens (Walbaum, 1792) NT MINV SB R RA SWA Rhinodaros percellens (Walbaum, 1784) VJ PLANK WC RA SWA/FA Sphirna devinit (Griffith & Smith, 1834) E MCAR WC RA CG Sphyrna telurius (Inmaeus, 1758) LC HERB RS/INIT/SB R CG Acanthurus coeru	Lamnidae	Isurus oxyrhincus	Rafinesque, 1810	NO	MCAR	WC	R	A	CG	LIT
Manta biostris (Walbaum, 1792) NT PLANK WC UN CG Mobula thurstoni (Lloyd, 1908) NT PLANK WC UN CG Nachula thurstoni (Lloyd, 1908) NT PLANK WC WA CG Nachula thurstoni (Niller & Henle, 1841) VU MCAR WC/RS RA CG Rhinobatos paccellens (Walbaum, 1792) NT MINV SB RA CG Rhinobatos paccellens (Walbaum, 1792) NT MINV SB RA CG Rhinocdon typus Smith, 1828 VU PLANK WC RA CT Sphyma devini (Griffith & Smith, 1834) LC MCAR WC RA CG Sphyma devini (Linneaus, 1758) LC MCAR WC RA CG Sphyma devini (Linneaus, 1758) LC HERB RS/INT/SB R CG Acanthurus chirurus (Bloch, 1787) (Bloch, 1787) LC H	Myliobatidae	Aetobatus narinari	(Euphrasen, 1790)	IN	MINV	WC		A	ل	SIG/LIT/PHO ^{1,2,3}
Mobula thurstoni (Lloyd, 1908) NT PLANK WC UN CG Narcine brasiliensis (Olfers, 1831) DD MINV SB R OC WA Carcharios taurus Rafinesque, 1810 VU MCAR WCARS RA CG Rhinobatos brokelir Muller & Henle, 1841 NT MINV SB RA CG Rhinobatos percellens (Walbaum, 1792) NT MINV SB RA CG Zapteryx brevirostris (Wuller & Henle, 1841) VU MINV SB R RA SWAFA Sphyma lewini (Griffith & Smith, 1828) VU PLANK WC RA CT Sphyma lewini (Griffith & Smith, 1834) EN MCAR WC RA CG Sphyma lewini (Griffith & Smith, 1834) EN MCAR WC RA CG Sphyma lewini (Linnaeus, 1758) LC MCAR WC RA CG Acanthurus balianus (Bloch & Schneid		Manta birostris	(Walbaum, 1792)	ΙΝ	PLANK	WC	Э	Z	CG	П
Avactione brasiliensis Olfers, 1831) DD MINV SB R OC WA Carcharias taurus Rafinesque, 1810 VU MCAR WC/RS RA CG Rhinobatos horkelii Muller & Henle, 1841 CR MINV SB RA SWA Rhinobatos percellens (Walbaum, 1792) NT MINV SB RA SWA/FA Aphinobatos percellens (Wuller & Henle, 1841) VU MINV SB R RA SWA/FA Aphinodou typus Smith, 1828 VU PLANK WC RA CT Sphyma lewini (Griffith & Smith, 1834) EN MCAR WC RA CG Sphyma lewini (Linnaeus, 1758) LC MCAR WC RA CG Sphyma lewini (Linnaeus, 1758) LC MCAR WC RA CG Sphyma sygaena (Linnaeus, 1758) LC HERB RS/INT/SB R C AO Acanthurus berliugus (Mobula thurstoni	(Lloyd, 1908)	TN	PLANK	WC	Π	N	CG	LIT
Carcharias taurus Rafinesque, 1810 VU MCAR WC/RS RA CG Rhinobatos horkelir Muller & Henle, 1841 CR MINV SB RA SWA Rhinobatos percellens (Walbaum, 1792) NT MINV SB RA SWAFA Zapteryx brevirostris (Muller & Henle, 1841) VU MINV SB RA SWAFA Sphyrra lewini (Griffith & Smith, 1834) EN MCAR WC RA CT Sphyrra lewini (Linnaeus, 1758) LC MCAR WC RA CG Acanthurus bahianus+ Castelnau, 1855 LC HERB RS/INT/SB R CG Acanthurus coeruleus* Bloch & Schneider, 1801 L HERB RS/INT/SB R RA Acanthurus monroviae* Steindachner, 1876 L PLANK WC/RS/INT R R	Narcinidae	Narcine brasiliensis	(Olfers, 1831)	DD	MINV	SB		C	WA	SIG/LIT/VOU/PHO ^{1,2,3}
Rhinobatos horkelii Muller & Henle, 1841 CR MINV SB RA SWA/EA Rhinobatos percellens (Walbaum, 1792) NT MINV SB RA SWA/EA Zapteryx brevirostris (Muller & Henle, 1841) VU MINV SB R RA SWA/EA Sphyrna lewini (Griffith & Smith, 1834) EN MCAR WC RA CT Sphyrna lewini (Linnaeus, 1758) LC MCAR WC RA CG Sphyrna zygaena (Linnaeus, 1758) LC MCAR WC RA CG Acanthurus chirurus chirurus (Bloch, 1787) LC HERB RS/INT/SB R AO Acanthurus coeruleus* Bloch & Schneider, 1801 LC HERB RS/INT/SB R RA Acanthurus monroviae* Steindachner, 1876 LC PLANK WC/RS/INT RA RA	Odontaspididae	Carcharias taurus	Rafinesque, 1810	N	MCAR	WC/RS	R	A	CG	LIT/PHO⁴
Rhinobatos percellens (Walbaum, 1792) NT MINV SB RA SWA/EA Zapteryx brevirostris (Muller & Henle, 1841) VU MINV SB R RA SWA Rhincodon typus Smith, 1828 VU PLANK WC RA CT Sphyrna lewini (Griffith & Smith, 1834) EN MCAR WC RA CG Sphyrna lewini (Linnaeus, 1758) LC MCAR WC RA CG Sphyrna zygaena (Linnaeus, 1758) LC HERB RS/INT/SB R CG Acanthurus chirurus chirurgus (Bloch, 1787) LC HERB RS/INT/SB R OC AO Acanthurus coeruleus* Bloch & Schneider, 1801 LC HERB RS/INT/SB R RA KA Acanthurus monroviae* Steindachner, 1876 LC PLANK WC/RS/INT RA KA	Rhinobatidae	Rhinobatos horkelii	Muller & Henle, 1841	CR	MINV	SB	R	A	SWA	SIG/LIT/PHO³
Zapteryx brevirostris (Muller & Henle, 1841) VU MINV SB RA SWA Rhincodon typus Smith, 1828 VU PLANK WC RA CT Sphyrna lewini (Griffith & Smith, 1834) EN MCAR WC RA CG Sphyrna lewini (Linnaeus, 1758) VU MCAR WC RA CG Acanthurus bahianus+ Castelnau, 1855 LC HERB RS/INT/SB R OC SWA/MAR Acanthurus coeruleus* Bloch & Schneider, 1801 LC HERB RS/INT/SB R OC AO Acanthurus monroviae* Steindachner, 1876 LC PLANK WC/RS/INT RA RA RA		Rhinobatos percellens	(Walbaum, 1792)	ΙΝ	MINV	SB	~	A	SWA/EA	LIT/VOU/PHO ³
Rhincodon typusSmith, 1828VUPLANKWCRACTSphyrna lewini(Griffith & Smith, 1834)ENMCARWCRACGSphyrna lewini(Linnaeus, 1758)LCMCARWCRAWA/TEPSphyrna zygaena(Linnaeus, 1758)LCHERBRS/INT/SBRCGSWA/MARAcanthurus bahianus+(Bloch, 1787)LCHERBRS/INT/SBROCAOAcanthurus coeruleus*Bloch & Schneider, 1801LCHERBRS/INT/SBRRAWA/MARAcanthurus monroviae*Steindachner, 1876LCPLANKWC/RS/INTRAEA		Zapteryx brevirostris	(Muller & Henle, 1841)	۸n	MINV	SB		А	SWA	LIT/VOU
Sphyrna lewini Grifffth & Smith, 1834) EN MCAR WC RA CG Sphyrna tiburo (Linnaeus, 1758) LC MCAR WC RA VA/TEP ae Acanthurus bahianus+ Castelnau, 1855 LC HERB RS/INT/SB R OC SWA/MAR Acanthurus coeruleus* Bloch & Schneider, 1801 LC HERB RS/INT/SB R OC AO Acanthurus monroviae* Steindachner, 1876 LC PLANK WC/RS/INT RA EA	Rhincodontidae	Rhincodon typus	Smith, 1828	۸n	PLANK	WC	R	А	CT	LIT/PHO
Sphyrna tiburo(Linnaeus, 1758)LCMCARWCRAWA/TEPSphyrna zygaena(Linnaeus, 1758)VUMCARWCRACGAcanthurus bahianus+Castelnau, 1855LCHERBRS/INT/SBROCAOAcanthurus coeruleus*Bloch & Schneider, 1801LCHERBRS/INT/SBROCAOAcanthurus monroviae*Steindachner, 1876LCPLANKWC/RS/INTRAEA	Sphyrnidae	Sphyrna lewini	(Griffith & Smith, 1834)	EN	MCAR	WC	~	A	CG	LIT/VOU
Sphyrna zygaena(Linnaeus, 1758)VUMCARWCRACGAcanthurus bahianus+Castelnau, 1855LCHERBRS/INT/SBROCSWA/MARAcanthurus coeruleus*Bloch & Schneider, 1801LCHERBRS/INT/SBROCAOAcanthurus monroviae*Steindachner, 1876LCPLANKWC/RS/INTRAEA		Sphyrna tiburo	(Linnaeus, 1758)	CC	MCAR	WC	~	A	WA/TEP	Ш
Acanthurus bahianus+Castelnau, 1855LCHERBRS/INT/SBROCSWA/MARAcanthurus coeruleus*Bloch & Schneider, 1801LCHERBRS/INT/SBRAOAcanthurus monroviae*Steindachner, 1876LCPLANKWC/RS/INTRAEA		Sphyrna zygaena	(Linnaeus, 1758)	NU	MCAR	WC	R	A	CG	VOU
(Bloch, 1787) LC HERB RS/INT/SB R OC AO * Steindachner, 1876 LC PLANK WC/RS/INT RA WA/MAR	Acanthuridae	Acanthurus bahianus+	Castelnau, 1855	CC	HERB	RS/INT/SB		Q	SWA/MAR	SIG/LIT/VOU/PHO ^{1,2,3}
Bloch & Schneider, 1801 LC HERB RS/INT/SB R RA WA/MAR * Steindachner, 1876 LC PLANK WC/RS/INT RA EA		Acanthurus chirurgus	(Bloch, 1787)	CC	HERB	RS/INT/SB		Q	AO	SIG/LIT/VOU/PHO ^{1,2,3}
Steindachner, 1876 LC PLANK WC/RS/INT RA EA		Acanthurus coeruleus*	Bloch & Schneider, 1801	CC	HERB	RS/INT/SB		V	WA/MAR	SIG/LIT/PHO ^{1,3}
		Acanthurus monroviae*	Steindachner, 1876	Ŋ	PLANK	WC/RS/INT	~	A	EA	PHO ³

Table 1. Continued.

Family	Species	Authority	IOCN	Trophic	Habitat	RE	Occur.	Geog. range	Rec.Type
Apogonidae	Apogon americanus*+	Castelnau, 1855	N N	PLANK	RS	Я	00	SWA	РНО³
	Apogon pseudomaculatus	Longley, 1932	N/N/N/N/N/N/N/N/N/N/N/N/N/N/N/N/N/N/N/	PLANK	RS	8	00	WA/EA	SIG/LIT/PHO ^{1,2,3}
	Phaeoptyx pigmentaria	(Poey, 1860)	Ŋ	PLANK	RS	æ	00	WA/EA	SIG/LIT/PHO ^{1,2,3}
Antennariidae	Antennarius striatus	(Shaw, 1794)	IJZ	MCAR	RS/INT	~	RA	b	ПТ
	Histrio histrio	(Linnaeus, 1758)	NE	MCAR	RS/INT		RA	C	ПТ
Ariidae	Cathorops spixii	(Agassiz, 1829)	NE	MCAR	SB		00	WA	VOU
	Genidens barbus	(Lacepède, 1803)	N/	MCAR	SB		90	SWA	NOU
	Genidens genidens	(Cuvier, 1829)	Ol	MCAR	RS/INT/SB		RA	SWA	SIG/LIT/VOU/PHO ^{1,3}
Balistidae	Balistes capriscus	Gmelin, 1789	NZ.	MINV	RS/INT/SB	~	00	WA/EA/MED	SIG/LIT/VOU
	Balistes vetula	Linnaeus, 1758	۸۸	MINV	RS/INT/SB	œ	RA	AO	SIG/LIT/PHO ^{1,3}
Batrachoididae	Porichthys porosissimus	(Cuvier, 1829)	N N	MCAR	RS/INT/SB	æ	8	SWA	SIG/LIT/VOU/PHO ^{2,3}
	Thalassophryne montevidensis	(Berg, 1893)	NE	MCAR	SB/INT		RA	SWA	ПТ
Belonidae	Strongylura marina	(Walbaum, 1792)	NE	OMNI	WC		00	WA	vou
	Tylosurus acus	(Lacepède, 1803)	N	MCAR	WC		RA	Cl	ПТ
Blenniidae	Hypleurochilus fissicornis	(Quoy & Gaimard, 1824)	N N	MINV	RS/INT	æ	8	SWA/EA	SIG/LIT/VOU/PHO ^{1,3}
	Hypleurochilus pseudoaequipinnis	Bath, 1994	NZ	MINV	RS/INT	~	00	WA	LIT/VOU
	Hypsoblennius invemar	Smith-Vaniz & Acero, 1980	N N	MINV	RS	8	8	WA	SIG/LIT/VOU
	Ophioblennius trinitatis+	Miranda-Ribeiro, 1919	NZ	HERB	RS	~	8	SWA	SIG/LIT/VOU/PHO1.3
	Parablennius marmoreus	(Poey, 1876)	NZ	OMNI	RS/INT	~	8	WA	SIG/LIT/VOU
	Parablennius pilicornis	(Cuvier, 1829)	N N	OMNI	RS/INT	~	8	SWA/EA/MED/WIO	SIG/LIT/VOU
	Scartella cristata	(Linnaeus, 1758)	NE	HERB	RS	Я	00	WA/EA/NWP	SIG/LIT/VOU/PHO ^{2,3}
Bothidae	Bothus ocellatus	(Agassiz, 1831)	NE NE	MINV	SB	Я	00	WA	SIG/LIT/VOU
	Bothus maculiferus	(Poey, 1860)	N N	MINV	SB		00	WA/EA	SIG/LIT/PHO ³
Callionymidae	Callionymus bairdi	Jordan, 1888	NE	MINV	RS/INT/SB	æ	RA	AO	LIT/VOU/PHO ³
Carangidae	Alectis ciliaris	(Bloch, 1787)	C	MCAR	WC		RA	Cl	SIG/LIT/PHO ³
	Caranx crysos	(Mitchill, 1815)	Ŋ	MCAR	WC	œ	00	AO	SIG/LIT/VOU/PHO ³
	Caranx hippos	(Linnaeus, 1766)	N	MCAR	WC	<u>«</u>	RA	WA/EA	SIG/LIT/VOU/PHO ³
	Caranx latus	Agassiz, 1831	NE	MCAR	WC	œ	RA	AO	SIG/LIT/VOU/PHO ^{1,3}
	Chloroscombrus chrysurus	(Linnaeus, 1766)	NE	PLANK	WC	<u>«</u>	RA	WA/EA	LIT/VOU
	Decapterus macarellus	(Cuvier, 1833)	Z	PLANK	WC		00	CG	SIG/LIT/PHO ³
	Decapterus punctatus*	(Cuvier, 1829)	N N	PLANK	WC		00	AO	SIG/PHO ³
	Naucrates ductor	(Linnaeus, 1758)	NE	MCAR	WC		RA	b	Ш
	Oligoplites saliens	(Bloch, 1793)	NE	PLANK	WC		RA	WA	SIG/LIT/VOU/PHO ³
	Oligoplites saurus	(Bloch & Schneider, 1801)	N.	MCAR	WC		RA	WA/TEP	LIT/VOU
	Pseudocaranx dentex	(Bloch & Schneider, 1801)	N N	PLANK	WC/SB	œ	8	Ь	SIG/LIT/VOU
	Selene setapinnis	(Mitchill, 1815)	Ŋ.	MCAR	WC		RA	WA	VOU
	Selene vomer	(Linnaeus, 1758)	IJ.	MCAR	WC		00	WA	SIG/LIT/VOU/PHO ³
	Seriola dumerili	(Risso, 1810)	E N	MCAR	WC	æ	00	9)	SIG/LIT/PHO ^{1,2,3}
	Seriola lalandi	Valenciennes, 1833	IJ.	MCAR	WC	œ	00	CG	SIG/LIT/PHO ^{1,3}
	Seriola rivoliana	Valenciennes, 1833	N N	MCAR	WC	8	00	CG	SIG/LIT/PHO ^{1,2,3}
	Trachinotus carolinus	(Linnaeus, 1766)	NE	MCAR	WC	~	00	WA	SIG/LIT/VOU/PHO ³
	Trachinotus falcatus	(Linnaeus, 1758)	N N	MCAR	WC	~	00	WA	SIG/LIT/VOU/PHO ^{1,3}
	Trachinotus goodei	Jordan & Evermann, 1896	NE	MCAR	WC	R	OC	WA	SIG/LIT/PHO ^{1,3}
									Continued

 Table 1. Continued.

Family	Species	Authority	NUO NUO NUO NUO NUO Nuo Nuo Nuo Nuo Nuo Nuo Nuo Nuo Nuo Nuo	Trophic	Habitat	뿚	Occur.	Geog. range	Kec. I ype
	Trachinotus marginatus	(Cuvier, 1832)	NE.	MCAR	WC		8	WA	SIG/LIT/VOU/PHO ³
Centropomidae	Centropomus undecimalis	(Bloch: 1792)	N.	MCAR	RS/INT/SB	8	9	WA	SIG/LIT/YOU
	Centropomus parallelus	Poey, 1860	N N	MCAR	RS/INT/SB		8 8	WA	SIG/LIT/VOU/PHO ³
Chaenopsidae	Emblemariopsis signifer	(Ginsburg, 1942)	C	MINV	RS/INT	R	9	WA	SIG/LIT/VOU
Chaetodontidae	Chaetodon sedentarius*	Poey, 1860	CC	SINV	RS/INT/SB		RA	WA/EA	LIT/PHO ³
	Chaetodon striatus	Linnaeus, 1758	CC	SINV	RS/INT/SB	R	9	WA	SIG/LIT/VOU
	Prognathodes guyanensis	(Durand, 1960)	C	SINV	RS/INT/SB	W.	RA	WA	LIT/VOU
Cirrhitidae	Amblycirrhitus pinos	(Mowbray, 1927)	N.	MINV	RS/INT	A.	RA	WA/MAR	LIT/PHO ^{1,2,3}
Clupeidae	Harengula clupeola	(Cuvier, 1829)	Ŋ	PLANK	WC		9	WA	VOU/PHO ^{1,3}
	Opisthonema oglinum	(LeSueur, 1818)	Ŋ	PLANK	WC		RA	WA	LIT/VOU
	Sardinella aurita	Valenciennes, 1847	Ŋ	PLANK	WC	_	RA	WA/EA	Ш
	Sardinella brasiliensis	(Steindachner, 1879)	NE.	PLANK	WC	Ū	9	WA	SIG/LIT/VOU/PHO3
Dactylopteridae	Dactylopterus volitans	(Linnaeus, 1758)	NE	MINV	INT/SB	R (OC	WA/EA	SIG/LIT/VOU
Dactyloscopidae	Dactyloscopus crossotus	Starks, 1913	NE.	MINV	INT/SB		RA	WA	LIT/VOU
	Dactyloscopus foraminosus	Dawson, 1982	C	MINV	INT/SB		RA	WA	늬
	Dactyloscopus tridigitatus	Gill, 1859	C	MINV	INT/SB		RA	WA	Ш
Diodontidae	Chilomycterus reticulatus	(Linnaeus, 1758)	N.	SINV	RS/INT/SB	R	RA	9)	SIG/LIT/VOU/PHO
	Chilomycterus spinosus	(Linnaeus, 1758)	빙	SINV	RS/INT/SB	W.	00	SWA	SIG/LIT/VOU/PHO1,3
	Diodon holocanthus	Linnaeus, 1758	N	SINV	RS/INT/SB		00	Ь	SIG/LIT/PHO ³
	Diodon hystrix	Linnaeus, 1758	NE	SINV	RS/INT/SB	R (OC	CT	SIG/LIT/PHO ^{1,3}
Echeneidae	Echeneis naucrates	Linnaeus, 1758	NE	MCAR	WC		RA	כן	LIT/VOU/PHO ^{2,3}
	Remora remora	(Linnaeus, 1758)	NE	MCAR	WC	_	RA	CT	LIT
Eleotridae	Eleotris pisonis	(Gmelin, 1789)	NE	MINV	RS/INT	1	RA	WA	LIT/VOU
Ephippidae	Chaetodipterus faber	(Broussonet, 1782)	NE	MINV	WC	R	0)	WA	SIG/LIT/VOU
Epinephelidae	Epinephelus adscensionis	(Osbeck, 1765)	CC	MCAR	RS/INT/SB	A.	RA	AO	SIG/LIT
	Epinephelus itajara	(Lichtenstein, 1822)	CR	MCAR	RS/INT/SB	æ	RA	WA/EA	LIT/PHO ^{1,2,3}
	Epinephelus marginatus	(Lowe, 1834)	EN	MCAR	RS/INT/SB	R	9	WA/EA	SIG/LIT/VOU
	Epinephelus morio	(Valenciennes, 1828)	Ä	MCAR	RS/INT/SB	R	00	WA	SIG/LIT/PHO ^{1,3}
	Hyporthodus niveatus	(Valenciennes, 1828)	N	MCAR	RS/INT/SB	R	9	WA	SIG/LIT/VOU
	Mycteroperca acutirostris	(Valenciennes, 1828)	CC	MCAR	RS/INT/SB	R	9	WA	SIG/LIT/VOU/PHO1,2,3
	Mycteroperca bonaci	(Poey, 1860)	Ā	MCAR	RS/INT/SB	R	00	WA	SIG/LIT/VOU/PHO1,2,3
	Mycteroperca interstitialis	(Poey, 1860)	N	MCAR	RS/INT/SB	W.	RA	WA	SIG/LIT/PHO ^{1,3}
	<i>Mycteroperca microlepis</i>	(Goode & Bean, 1879)	C	MCAR	RS/INT/SB	ж Б	RA	WA	SIG/LIT/PHO3
	Paranthias furcifer	(Valenciennes, 1828)	C	PLANK	INT/SB		RA	WA/EA	SIG/LIT/PHO ^{1,3}
Fistulariidae	Fistularia petimba	Lacepède, 1803	NE	MCAR	RS/INT/SB		RA	WA/EA/MED/IP	NON
	Fistularia tabacaria	Linnaeus, 1758	NE	MCAR	RS/INT/SB	R (CO	WA/EA	SIG/LIT/VOU/PHO ^{1,3}
Gerreidae	Diapterus auratus	Ranzani, 1842	NE	MINV	INT/SB)	0)	WA	SIG/LIT/VOU/PHO ³
	Diapterus rhombeus	(Cuvier, 1829)	Ŋ	MINV	INT/SB	J	00	WA	NON
	Eucinostomus argenteus	Baird & Girard, 1855	NE	MINV	INT/SB	J	00	EP/WA/EA	NON
	Eucinostomus gula	(Quoy & Gaimard, 1824)	N.	MINV	INT/SB	0	9	WA	SIG/LIT/VOU/PHO3
	Eucinostomus melanopterus	(Bleeker, 1863)	NE	MINV	INT/SB		9	WA/EA	SIG/LIT/VOU/PHO3
	Eugerres brasilianus	(Cuvier, 1830)	NE	MINV	INT/SB		00	WA	SIG/LIT/VOU/PHO ³
Gobiesocidae	Gobiesox barbatulus	Starks, 1913	N	MINV	RS		RA	WA	SIGNITA/OLI/BHO3

Table 1. Continued.

	٠	2							6	
ramııy	Species	Authority	IOCN	Iropnic	Habitat	KE O	Occur.	Geog. range	Kec. Iype	
	Tomicodon australis	Briggs 1955	NE	MINV	RS	RA	A	WA	Non	
Gobiidae	Barbulifer ceuthoecus	(Jordan & Gilbert, 1884)	NE	MINV	INT/SB	RA	A	WA	LIT/VOU	
	Bathygobius soporator	(Valenciennes, 1837)	N	MINV	RS/INT/SB	8	0	EA/WA/MED	SIG/LIT/VOU/PHO ³	
	Coryphopterus glaucofraenum	Gill, 1863	N	OMNI	INT/SB	R CO	0	WA	SIG/LIT/VOU	
	Ctenogobius saepepallens	(Gilbert & Randall, 1968)	NE	MINV	INT/SB	RA	4	WA	SIG/LIT/PHO ^{1,3}	
	Ctenogobius stigmaticus	(Poey, 1860)	NE	MINV	RS/INT/SB	RA	δ	WA	LIT/VOU	
	Elacatinus figaro+	Sazima, Moura & Rosa, 1996	NE	MINV	RS/INT/SB	R RA	٨	SWA	LIT/VOU	
	Gnatholepis thompsoni	Jordan, 1904	N	MINV	RS/INT/SB	R RA	A	WA	SIG/LIT	
	Gobiosoma hemigymnum	(Eingenmann & Eingenmann, 1888)	NE	MINV	RS/INT/SB	RA	A	WA	LIT/PHO ³	
	Microgobius meeki	Evermann & Marsh, 1899	NE	MINV	RS/INT/SB	RA	V	WA	LIT/VOU	
Haemulidae	Anisotremus surinamensis	(Bloch, 1791)	NE	MINV	RS/INT	R CO	0	WA	SIG/LIT/VOU	
	Anisotremus virginicus	(Linnaeus, 1758)	NE	MINV	RS/INT	R CO	C	WA	SIG/LIT/VOU	
	Haemulon aurolineatum	Cuvier, 1830	NE	MINV	RS/INT/SB	R	C	WA	SIG/LIT/VOU	
	Haemulon steindachneri	(Jordan & Gilbert, 1882)	NE	MINV	RS/INT/SB	R 0C	U	WA	SIG/LIT/VOU/PHO ³	
	Haemulon parra	(Desmarest, 1823)	NE	MINV	RS/INT/SB	R 0C	U	WA	SIG/LIT/PHO ³	
	Orthopristis ruber	(Cuvier, 1830)	NE	MINV	RS/INT/SB	R CO	C	WA	SIG/LIT/VOU	
Hemiramphidae	Hemiramphus brasiliensis	(Linnaeus, 1758)	NE	OMNI	WC	00	U	WA/EA	SIG/LIT/VOU/PHO ³	
	Hyporhamphus unifasciatus	(Ranzani, 1841)	NE	OMNI	WC	RA	A	WA	SIG/LIT/VOU/PHO ³	
Holocentridae	Holocentrus adscensionis	(Osbeck, 1765)	NE	MINV	RS/INT/SB	R CO	0	AO	SIG/LIT/VOU	Ī
	Myripristis jacobus	Cuvier, 1829	NE	MINV	RS/INT	R RA	٨	AO	SIG/LIT/VOU	
Kyphosidae	Kyphosus sectatrix	(Linnaeus, 1758)	NE	HERB	RS	R CO	0	Cl	SIG/LIT/PHO ³	
	Kyphosus vaigiensis	(Quoy & Gaimard, 1825)	N	HERB	RS	R CO	C	Ь	SIG/LIT/VOU	
Labridae -	Bodianus pulchellus	(Poey, 1860)	C	MINV	RS/INT	R 0C	U	WA/EA	SIG/LIT/VOU	
Hypsigeninae	Bodianus rufus	(Linnaeus, 1758)	S	MINV	RS/INT	R CO	0	WA	SIG/LIT/PHO ^{1,2,3}	
	Clepticus brasiliensis*+	Heiser, Moura & Robertson, 2000	C	PLANK	RS/WC	R RA	A	SWA	SIG/LIT/PHO ^{1,3}	
Labridae - Julidinae	Doratonotus megalepis	Günther 1862	C	MINV	RS	R RA	A	WA	SIG/LIT	
	Halichoeres brasiliensis+	(Bloch, 1791)	Ŋ	MINV	RS/INT/SB	R 0C	U	SWA	SIG/LIT/PHO ^{1,3}	
	Halichoeres dimidiatus	(Agassiz, 1831)	Ŋ	MINV	RS/INT/SB	R RA	٨	SWA	SIG/LIT/PHO ³	
	Halichoeres poeyi	(Steindachner, 1867)	C	MINV	RS/INT/SB	R CO	0	WA	SIG/LIT/VOU	
	Halichoeres sazimai+	Luiz, Ferreira & Rocha, 2009	NE	MINV	RS/INT/SB	R RA	A	SWA	SIG/LIT/PHO ³	
	Thalassoma noronhanum+	(Boulenger, 1890)	Ŋ	PLANK	RS/INT/SB	R RA	δ	SWA	SIG/LIT/PHO ^{1,3}	
	<i>Xyrichtys novacula</i>	(Linnaeus, 1758)	C	MINV	SB	R RA	٨	WA/EA	SIG/LIT/VOU/PHO ^{1,3}	
Labridae - Scarini	Cryptotomus roseus	Cope, 1871	Ŋ	HERB	RS/INT/SB	R CO	0	WA/MAR	SIG/LIT/VOU/PHO ^{1,2,3}	
	Nicholsina usta	Valenciennes, 1840	Ŋ	HERB	RS/INT	R RA	δ	WA	SIG/LIT/PHO ³	
	Scarus trispinosus+	Valenciennes, 1840	EN	HERB	RS/INT/SB	R RA	٨	SWA	LIT/PHO ^{1,3}	
	Scarus zelindae+	Moura, Figueiredo & Sazima, 2001	OO	HERB	RS/INT/SB	R RA	٨	SWA	SIG/LIT/PHO ^{1,3}	
	Sparisoma amplum+	(Ranzani, 1841)	Ŋ	HERB	RS/INT/SB	R CO	0	SWA	SIG/LIT/PHO ^{1,2,3}	
	Sparisoma axillare+	(Steindachner, 1878)	Ŋ	HERB	RS/INT/SB	R CO	0	SWA	SIG/LIT/PHO ^{1,2,3}	
	Sparisoma frondosum+	(Agassiz, 1831)	DD	HERB	RS/INT/SB	R	0	SWA/EA	SIG/LIT/PHO ^{1,2,3}	
	Sparisoma radians	(Valenciennes, 1840)	Ŋ	HERB	RS/INT/SB	R CO	0	SWA	SIG/LIT/PHO ^{1,3}	
	Sparisoma tuiupiranga+	Gasparini, Joyeux & Floeter, 2003	C	HERB	RS/INT/SB	R 0C	U	SWA	SIG/LIT/PHO ^{1,2,3}	
Labrisomidae	Labrisomus cricota+	Sazima, Gasparini & Moura, 2002	NE	MINV	RS/INT	R RA	4	SWA	SIG/LIT/VOU/PHO ³	

Table 1. Continued.

	<u>Labrisomus nuchipinnis</u>				Habitat	4	כנת:			
		(Quoy & Gaimard, 1824)	Ŋ	MINV	RS/INT	R	9	WA/EA	SIG/LIT/VOU	
	Malacoctenus delalandii	(Valenciennes, 1836)	NE	MINV	RS/INT	~	8	WA	SIG/LIT/PHO ^{1,2,3}	
	Malacoctenus aff. triangulatus+		R	MINV	RS/INT	æ	RA	SWA	LIT/VOU	
	Paraclinus rubicundus	(Starks, 1913)	2	MINV	RS/INT	æ	RA	SWA	П	
	Paraclinus spectator+	Guimarães & Bacelar, 2002	N N	MINV	RS/INT	æ	00	SWA	SIG/LIT/PHO ^{1,3}	
	Starksia brasiliensis+	(Gilbert, 1900)	N	MINV	RS/INT	~	00	SWA	SIG/LIT/VOU/PHO3	
Lobotidae	Lobotes surinamensis	(Bloch, 1790)	빙	MCAR	RS/INT		RA	D D	LIT/PHO ³	
Lutjanidae	Lutjanus analis	(Cuvier, 1828)	ΛΛ	MCAR	RS/INT/SB	R	RA	WA	SIG/LIT/VOU/PHO ^{1,3}	
	Lutjanus cyanopterus	(Cuvier, 1828)	N	MCAR	RS/INT/SB	æ	RA	WA	SIG/LIT/PHO ³	
	Lutjanus jocu	(Bloch & Schneider, 1801)	빙	MCAR	RS/INT/SB	æ	RA	WA/MAR	SIG/LIT/VOU/PHO ^{1,3}	
	Rhomboplites aurorubens	(Cuvier, 1829)	NE	MCAR	RS/INT	R	OC	WA	SIG/LIT/PHO ^{1,3}	
Malacanthidae	Malacanthus plumieri	(Bloch, 1786)	NE	MCAR	RS/INT/SB	R	OC	WA/MAR	SIG/LIT/VOU/PHO ^{1,3}	
Microdesmidae	Ptereleotris randalli+	Gasparini, Rocha & Floeter, 2001	NE	MINV	SB	R	OC	SWA	SIG/LIT/PHO ^{1,3}	
Monacanthidae	Aluterus monoceros	(Linnaeus, 1758)	NE	OMNI	RS/INT/SB	R	00	IJ	SIG/LIT/PHO ^{1,2,3}	
	Aluterus scriptus	(Osbeck, 1765)	NE NE	OMNI	RS/INT/SB	æ	RA	Ь	PHO/LIT	
	Cantherhines macrocerus*	(Hollard, 1853)	N N	SINV	RS	æ	RA	WA/EA	PHO	
	Monacanthus ciliatus	(Mitchill, 1818)	Ŋ	OMNI	RS/INT/SB	æ	RA	WA	LIT/VOU	
	Stephanolepis hispidus	(Linnaeus, 1766)	NE NE	OMNI	RS/INT/SB	æ	8	WA/EA	SIG/LIT/VOU	
	Stephanolepis setifer	(Bennett, 1831)	NE	OMNI	RS/INT/SB	R	RA	WA	П	
Mugilidae	Mugil curema	Valenciennes, 1836	뷩	OMNI	INT/SB	æ	8	WA/EA/TEP	SIG/LIT/VOU	
	Mugil Iiza	Valenciennes, 1836	NE	OMNI	INT/SB		00	WA	SIG/LIT/VOU/PHO ³	
Mullidae	Pseudupeneus maculatus	(Bloch, 1793)	NE	MINV	RS/INT/SB	R	00	WA	SIG/LIT/VOU	
	Upeneus parvus	Poey, 1852	NE	MINV	RS/INT/SB		RA	WA	LIT/PHO	
Muraenidae	Echidna catenata	(Bloch, 1795)	NE	MCAR	RS	R	RA	WA/MAR	LIT/VOU	
	Gymnothorax funebris	Ranzani, 1839	NE	MCAR	RS	~	00	WA	SIG/LIT/PHO ^{1,2,3}	
	Gymnothorax moringa	(Cuvier,1829)	NE	MCAR	RS	æ	8	WA/MAR	SIG/LIT/VOU	
	Gymnothorax ocellatus	Agassiz, 1831	NE	MCAR	RS		RA	AO	NON	
	Gymnothorax vicinus*	(Castelnau, 1855)	NE	MCAR	RS	æ	00	TA	PHO ³	
	Muraena retifera*	Goode & Bean, 1882	NE	MCAR	INT/SB	R	RA	WA	PHO	
Ogcocephalidae	Ogcocephalus vespertilio	(Linnaeus, 1758)	NE	MINV	RS/INT/SB	R	CO	WA	SIG/LIT/VOU	
Ophichthidae	Ahlia egmontis	(Jordan, 1884)	NE	MINV	RS/INT/SB	~	RA	WA	Ш	
	Myrichthys breviceps	(Richardson, 1848)	NE	MINV	RS/INT	æ	00	WA	SIG/LIT/VOU	
	Myrichthys ocellatus	(LeSueur, 1825)	NE	MINV	RS/INT	~	8	WA	SIG/LIT/VOU	
	Myrophis punctatus	Lütken, 1852	뵘	MINV	RS/INT/SB		RA	WA	Ш	
	Herpetoichthys regius*	(Richardson, 1848)	N	MINV	INT/SB	æ	N	EA	SIG/PHO ³	
	Ophichthusophis	(Castelnau, 1855)	NE	MINV	INT/SB		RA	WA	PHO	
Ophidiidae	Genipterus brasiliensis	Regan, 1903	NE	MINV	SB/INT		RA	SWA	Ш	
	Ophidion holbrooki	Putnam, 1874	NE	MINV	SB/INT		RA	WA	LIT/VOU	
	Raneya brasiliensis	(Kaup, 1856)	NE	MINV	SB/INT		RA	SWA	П	
Ostraciidae	Acanthostracion polygonius	Poey, 1876	NE	OMNI	RS/INT/SB	R	00	WA	SIG/LIT/PHO ^{1,2,3}	
	Acanthostracion quadricornis	(Linnaeus, 1758)	Ŋ	OMNI	RS/INT/SB	æ	00	WA/EA	SIG/LIT/VOU	
	Lactophys trigonus	(Linnaeus, 1758)	NE	OMNI	RS/INT/SB		RA	WA	LIT/VOU	
Paralichthyidae	Cyclopsetta fimbriata	(Goode & Bean, 1885)	NE	MINV	SB		OC	WA	SIG/LIT/PHO ³	

Table 1. Continued.

:										
Family	Species	Authority	IOCN	Trophic	Habitat	æ	Occur.	Geog. range	Rec.Type	
	Paralichthys brasiliensis	(Ranzani, 1842)	빙	MIN/	SB		00	SWA	LIT/VOU	
	Syacium micrurum	Ranzani, 1842	NE	MINV	SB		RA	WA	NON	
	Syacium papillosum	(Linnaeus, 1758)	NE	MINV	SB		RA	WA/MAR	LIT	
Pempheridae	Pempheris schomburgki	Müller & Troschel, 1848	NE	PLANK	RS/INT	R (CO	WA	SIG/LIT/VOU	
Polynemidae	Polydactylus oligodon	(Günther, 1860)	NE	OMNI	INT/SB		RA	WA	רוב	
	Polydactylus virginicus	(Linnaeus, 1758)	NE	OMNI	INT/SB		RA	WA	LIT/VOU	
Pomacanthidae	Centropyge aurantanotus	Burgess, 1974	CC	HERB	RS/INT	R	RA	WA/EA	SIG/LIT/PHO³	
	Holacanthus ciliaris	(Linnaeus, 1758)	Ŋ	SINV	RS/INT/SB	~	RA	WA	SIG/LIT/VOU	
	Holacanthus tricolor	(Bloch, 1795)	Ŋ	SINV	RS/INT/SB	- W	RA	WA	SIG/LIT/PHO ^{1,2,3}	
	Pomacanthus arcuatus	(Linnaeus, 1758)	C	SINV	RS/INT/SB	- -	RA	WA	SIG/LIT/PHO ^{1,3}	
	Pomacanthus paru	(Bloch, 1787)	C	SINV	RS/INT/SB	R	9	WA/MAR	SIG/LIT/VOU	
Pomacentridae	Abudefduf saxatilis	(Linnaeus, 1758)	핑	OMNI	RS/INT/SB	R	9	AO	SIG/LIT/VOU	
	Chromis flavicauda*	(Günther, 1880)	DD	PLANK	RS/INT	~	RA	WA/EA	SIG/LIT/PHO ³	
	Chromis jubauna+	Moura, 1995	N	PLANK	RS/INT	~	RA	SWA	SIG/LIT/VOU/PHO ^{1,3}	
	Chromis limbata	(Valenciennes, 1833)	N	PLANK	RS/INT	8	8	WA/EA	SIG/LIT/PHO ^{1,3}	
	Chromis multilineata	(Guichenot, 1853)	N	PLANK	RS/INT	~	9	AO	SIG/LIT/VOU	
	Stegastes fuscus+	(Cuvier, 1830)	Ŋ	HERB	RS/INT	8	9	SWA	SIG/LIT/VOU	
	Stegastes partitus*	(Poey, 1868)	N	HERB	RS/INT	W W	NO	NWA	SIG/PHO ³	
	Stegastes pictus+	(Castelnau, 1855)	빙	HERB	RS/INT	8	9	SWA	SIG/LIT/VOU	
	Stegastes variabilis	(Castelnau, 1855)	NE	HERB	RS/INT	R	CO	SWA	SIG/LIT/VOU	
Pomatomidae	Pomatomus saltatrix	(Linnaeus, 1766)	NE	MCAR	WC	R (00	90	SIG/LIT/VOU	
Priacanthidae	Cookeolus japonicus	(Cuvier, 1829)	NE	MINV	RS/INT	R	RA	CG	LIT/VOU	
	Priacanthus arenatus	Cuvier, 1829	NE	MINV	RS/INT	R	OC	WA/EA	SIG/LIT/VOU/PHO ^{1,3}	
Rachycentridae	Rachycentron canadum	(Linnaeus 1766)	NE	MCAR	RS/INT/SB		RA	CT	LIT/VOU	
Sciaenidae	Menticirrhus americanus	(Linnaeus, 1758)	NE	MINV	INT/SB		0)	WA	LIT/VOU/PHO	
	Menticirrhus littoralis	(Holbrook, 1847)	묑	MINV	INT/SB		00	WA	NOU	
	Micropogonias furnieri	(Desmarest, 1823)	묑	MINV	INT/SB		9	WA	LIT/VOU/PHO	
	Odontoscion dentex	(Cuvier, 1830)	R	MCAR	RS/INT	æ	8	WA	SIG/LIT/VOU	
	Pareques acuminatus	(Bloch & Schneider, 1801)	NE	MINV	RS/INT/SB	R	CO	WA	SIG/LIT/VOU	
Scombridae	Acanthocybium solandri	(Cuvier, 1832)	C	MCAR	WC		00	ט	5	
	Euthynnus alletteratus	(Rafinesque, 1810)	C	MCAR	WC		00	AO	LIT/PHO³	
	Scomberomorus brasiliensis	Collette, Russo & Zavala-Camin, 1978	Ŋ	MCAR	WC		00	WA	LIT/VOU	
	Scomberomorus cavalla	(Cuvier, 1829)	CC	MCAR	WC		OC	WA/EA	LIT	
Scorpaenidae	Scorpaena brasiliensis	Cuvier, 1829	NE	MCAR	RS/INT/SB	R	00	WA	SIG/LIT/PHO ^{1,3}	
	Scorpaena isthmensis	Meek & Hildebrand, 1928	R	MCAR	RS/INT/SB		00	WA	NOU	
	Scorpaena plumieri	Bloch, 1789	NE	MCAR	RS/INT/SB	R (OC	WA/MAR	SIG/LIT/VOU/PHO ^{1,3}	
Serranidae	Diplectrum formosum	(Linnaeus, 1766)	NE	MCAR	SB	R	0)	WA	SIG/LIT/VOU/PHO ^{1,3}	
	Diplectrum radiale	(Quoy & Gaimard, 1824)	N	MCAR	SB	R	8	WA	SIG/LIT/VOU/PHO ^{1,3}	
	Dules auriga	Cuvier, 1829	NE	MCAR	INT/SB		9	SWA	SIG/LIT/PHO ^{1,2,3}	
	Rypticus randalli	Courtenay, 1967	묑	MCAR	RS/INT		RA	WA/EA	LIT/VOU	
	Rypticus saponaceus	(Bloch & Schneider, 1801)	NE	MCAR	RS/INT	- R	RA	AO	ГП	
	Serranus atrobranchus	(Cuvier, 1829)	NE	MINV	INT/SB	R	9	WA	SIG/LIT/PHO ^{1,2,3}	
	Serranus baldwini	(Evermann & Marsch, 1899)	NE	MINV	INT/SB		CO	WA	SIG/LIT/PHO ^{1,2,3}	
									Continued	inued

Table 1. Continued.

Family	Species	Authority	INCN	Trophic	Habitat	RE	Occur.	Geog. range	Kec. I ype
	Serranus flaviventris	(Cuvier, 1829)	NE	MINV	INT/SB	R	00	WA	SIG/LIT/VOU
Sparidae	Archosargus probatocephalus	(Walbaum, 1792)	NE	MINV	RS/INT	В	00	WA	ПТ
	Archosargus rhomboidalis	(Linnaeus, 1758)	N	OMNI	RS/INT/SB	œ	RA	WA	SIG/LIT/VOU/PHO ³
	Calamus penna	(Valenciennes, 1830)	N	MINV	INT/SB	œ	RA	WA	SIG/LIT/PHO ^{1,3}
	Calamus pennatula	Guichenot, 1868	NE	MINV	INT/SB	œ	RA	WA	SIG/LIT/VOU/PHO ³
	Diplodus argenteus	(Valenciennes, 1830)	NE.	OMNI	RS/INT/SB	æ	9	SWA	SIG/LIT/VOU
	Pagrus pagrus	(Linnaeus, 1758)	EN	MCAR	RS/INT/SB		RA	WA	LIT/VOU
Sphyraenidae	Sphyraena barracuda	(Edwards, 1771)	NE	MCAR	RS/INT/SB	R	RA	CT	SIG/LIT/PHO ^{2,3}
	Sphyraena guachancho	Cuvier, 1829	NE NE	MCAR	RS/INT/SB	æ	00	WA/EA	SIG/LIT/VOU/PHO3
	Sphyraena tome	Fowler, 1903	NE	MCAR	RS/INT/SB	æ	00	SWA	NON
Syngnathidae	Halicampus crinitus	(Jenyns, 1842)	NE	MINV	RS/INT	R	00	WA	SIG/LIT/PHO ^{1,2,3}
	Hippocampus erectus	Perry, 1810	N	MINV	RS/INT	œ	00	WA/MAR	SIG/LIT/VOU
	Hippocampus patagonicus	Piacentino & Luzzato, 2004	N	MINV	RS/INT	æ	00	SWA	SIG/LIT/VOU/PHO3
	Hippocampus reidi	Ginsburg, 1933	NN	MINV	RS/INT	æ	00	WA	SIG/LIT/VOU
	Microphis lineatus	(Kaup, 1856)	JE N	MINV	RS/INT		RA	WA	NOU
	Syngnathus folletti	Herald, 1942	NE	MINV	RS/INT		RA	SWA	NOU
Synodontidae	Synodus intermedius	(Spix & Agassiz, 1829)	R	MCAR	RS/INT/SB	œ	RA	WA	LIT/PHO ³
	Synodus bondi	Fowler, 1939	NE.	MCAR	INT/SB		RA	WA	LIT/VOU/PHO3
	Synodus synodus	(Linnaeus, 1758)	J.	MCAR	RS/INT/SB	æ	9	AO	SIG/LIT/PHO ^{1,2,3}
	Trachynocephalus myops	(Forster, 1801)	DD	MCAR	INT/SB		RA	AO	LIT
Tetraodontidae	Canthigaster figueiredoi+	Moura & Castro, 2002	NE	OMNI	RS/INT/SB	R	00	WA	SIG/LIT/VOU
	Lagocephalus laevigatus	(Linnaeus, 1766)	R	OMNI	SB		00	WA/EA	LIT/VOU
	Sphoeroides greeleyi	Gilbert, 1900	NE.	MINV	RS/INT/SB	æ	8	WA	NOU
	Sphoeroides pachygaster	(Müller & Troschel, 1848)	NE.	MINV	RS/INT/SB		RA	9)	П
	Sphoeroides spengleri	(Bloch, 1785)	J.	MINV	RS/INT/SB	œ	9	WA/EA	SIG/LIT/VOU
	Sphoeroides testudineus	(Linnaeus, 1758)	JE N	MINV	RS/INT/SB	æ	8	WA	SIG/LIT/VOU
	Sphoeroides tyleri	Shipp, 1972	NE	MINV	RS/INT/SB		RA	WA	LIT/VOU
Triglidae	Prionotus nudigula	Ginsburg, 1950	NE	MINV	INT/SB		RA	SWA	NON
	Prionotus punctatus	(Bloch, 1793)	NE	MINV	INT/SB		RA	WA	LIT/VOU
Uranoscopidae	Astroscopus y-graecum	(Cuvier, 1829)	QQ	MCAR	INT/SB		RA	WA	LIT/PHO ³

Photographic records: 1. Anderson et al. 2014a 2. Hostim-Silva et al. 2006 3. Marine Macroecology and Biogeography Lab, Photographic Databank 4. Souza 2000

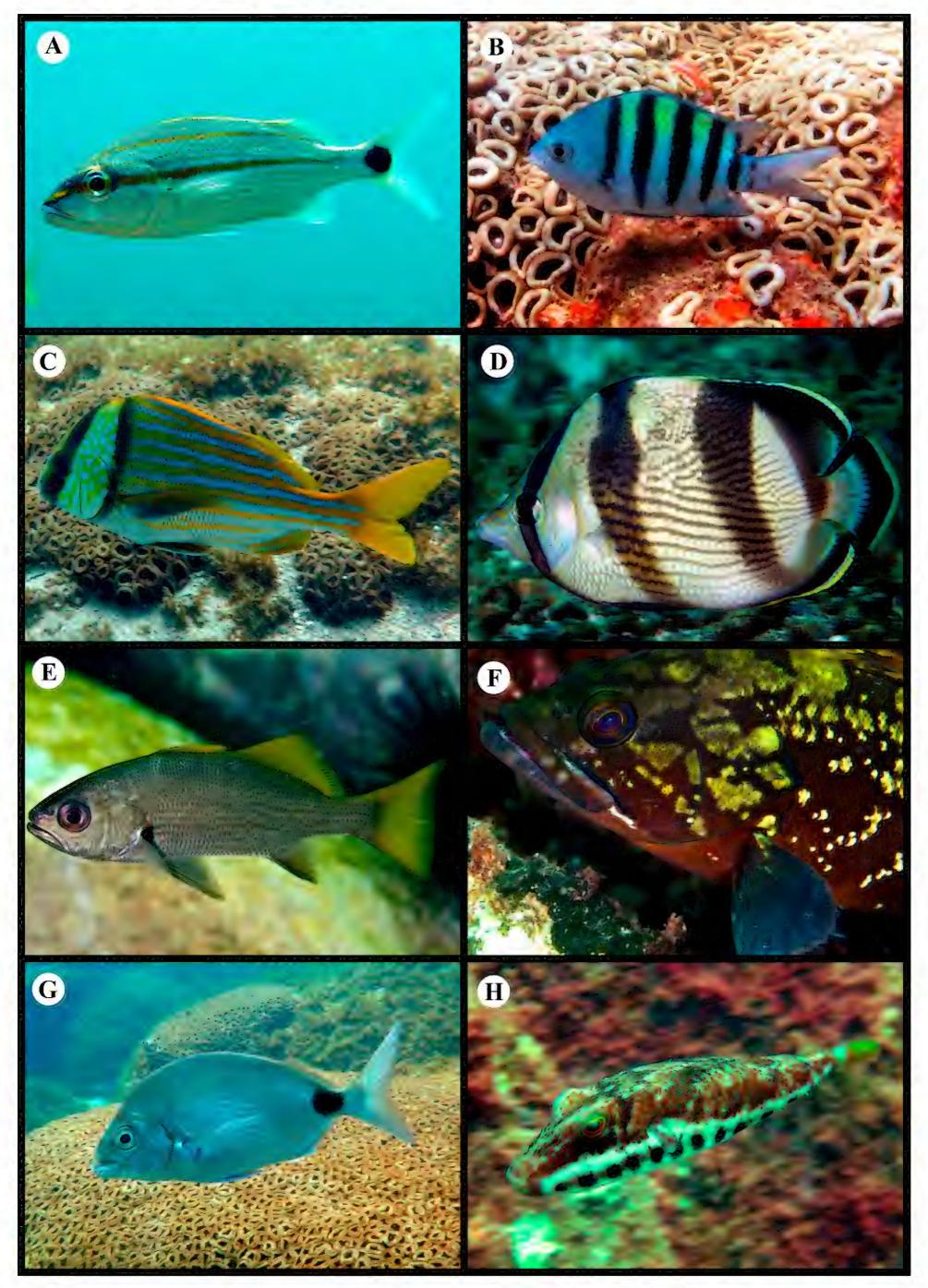


Figure 3. Examples reef fish considered "common" in reefs of Santa Catarina, Southern Brazil. (**A**) The Tomtate Grunt, *Haemulon aurolineatum*; (**B**) Sergeant Major, *Abudefduf saxatilis*; (**C**) Porkfish, *Anisotremus virginicus*; (**D**) Banded Butterflyfish, *Chaetodon striatus*; (**E**) Reef Croaker, *Odontoscion dentex*; (**F**) Dusky Grouper, *Epinephelus marginatus*; (**G**) South American Silver Porgy, *Diplodus argenteus*; (**H**) Bandtail Puffer, *Sphoeroides spengleri*.



Figure 4. Examples of reef fish species considered as "rare" in reefs of Santa Catarina, Southern Brazil. (**A**) Emerald Parrotfish, *Nicholsina usta*; (**B**) Guri Sea Catfish, *Genidens genidens*; (**C**) Creole-fish, *Paranthias furcifer*; (**D**) Sheepshead Porgy, *Calamus penna*; (**E**) Brazilian Guitarfish, *Rhinobatos horkelii*; (**F**) Queen Triggerfish, *Balistes vetula*; (**G**) Flameback Angelfish, *Centropyge aurantonotus*; (**H**) Honeycomb Cowfish, *Acanthostracion polygonius*.

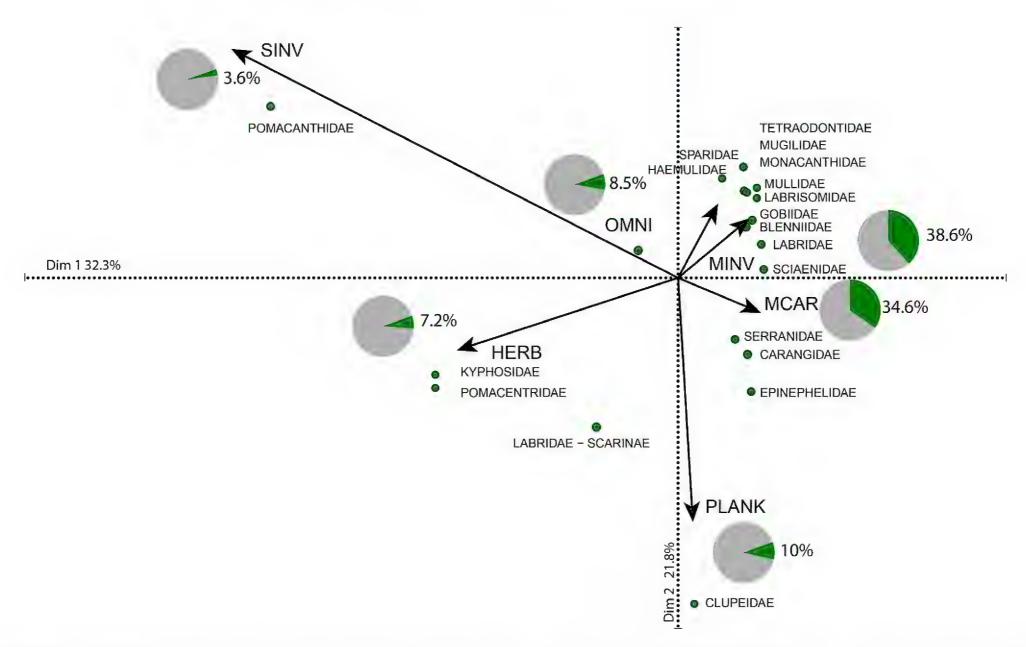


Figure 5. Correspondence analysis based on species richness showing the trophic structure of reef fish ichthyofauna in Santa Catarina's rocky reef systems. Vectors indicate the feeding habits of species: MINV = mobile invertebrate feeders, MCAR = macrocarnivores, PLANK = planktivores, OMNI = omnivores, HERB = herbivores/detritivores, SINV = sessile invertebrate feeders. Green circles mark the positions of the families. Grey/green circles represent the relative proportion of species for each trophic group.

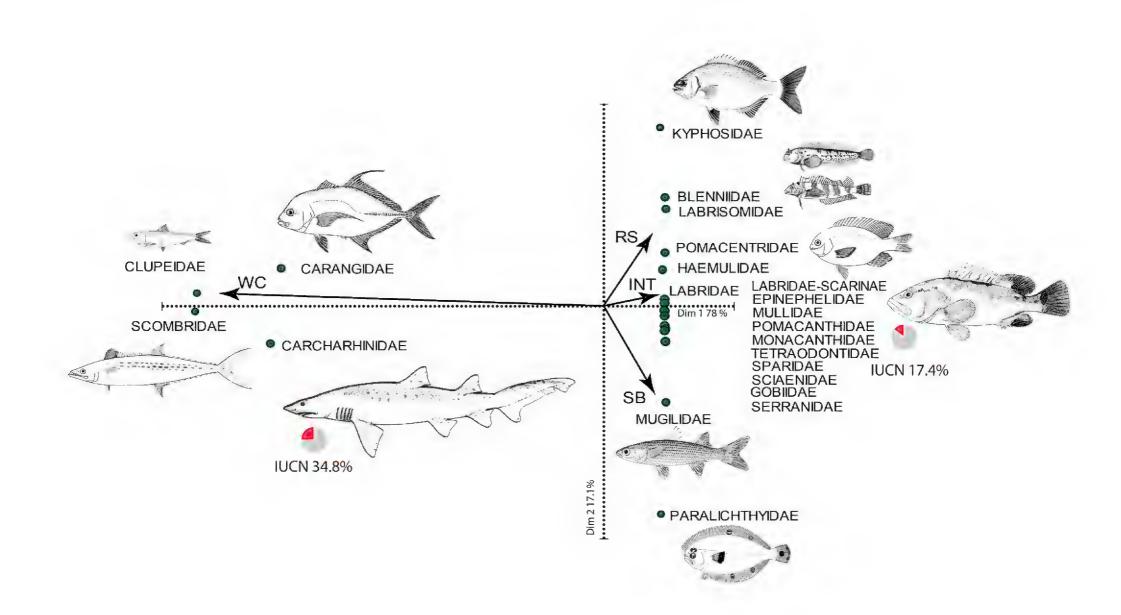


Figure 6. Habitat distribution of reef fish in Santa Catarina's rocky reef systems. The vectors indicate the reef's ecological zones: RS = Reef Slope zone, INT = interface, SB = sandy bottom, WC = water column. Green circles indicate the position of respective families in the rocky reef system (CA). Grey/red circles indicate the relative quantity of threatened species within the families of sharks, tunas and groupers, which are the most threatened groups.

americanus (Castelnau 1855); Cantherhines macrocerus (Hollard, 1853); Chaetodon sedentarius (Poeyi, 1860); Chromis flavicauda (Günther, 1880); Clepticus brasiliensis (Heiser, Moura & Robertson, 2000); Decapterus punctatus (Cuvier, 1829); Gymnothorax vicinus (Castelnau, 1855); Herpetoichthys regius (Richardson, 1848); Muraena retifera Goode & Bean, 1882 and Stegastes partitus (Poey, 1868). For S. partitus, this is the first documented record for Southwestern Atlantic waters, and for H. regius this is the first record for the coastal Southwestern Atlantic (Figure 7). For Acanthurus monroviae Steindachner, 1876, this is also the second record for Southwestern Atlantic waters. Details on the new records are given below (ordered alphabetically by Order and then Family).

Order Anguilliformes, Family Muraenidae

Gymnothorax vicinus (Castelnau, 1855) (Figure 7A). One individual was recorded at the approximate depth of 7 m, Arvoredo Marine Biological Reserve, Santa Catarina in 2008. *Remarks:* Previous southernmost record was the state of Paraná (Hackradt and Félix-Hackradt 2009).

Muraena retifera Goode & Bean, 1882 (Figure 7B). One adult individual was recorded at Xavier Island and another one at Aranhas Island in February 2015 both at the approximate depth of 8 m. *Remarks:* Previous southernmost record was the state of São Paulo (Carvalho-Filho 1999).

Order Anguilliformes, Family Ophichthidae

Herpetoichthys regius (Richardson, 1848) (Figure 7C and 7D). One large individual (around 90 cm) was recorded at Rancho Norte, Arvoredo Marine Biological Reserve in February 2014, at the approximate depth of 7 m, and another one smaller (around 40 cm) was recorded from Deserta Island, Arvoredo Marine Biological Reserve in May 2014. These two individuals were observed patrolling rhodolith banks that exist at these two locations. Remarks: These individuals represent the first record for the coastal Western Atlantic Ocean. Its native geographic range is the Eastern Atlantic, Ascension and Santa Helena Islands, but it has been recently reported to occur also on Saint Peter and Saint Paul Archipelago (Wirtzet al. 2015). Therefore this is the second report of this species for the Southwestern Atlantic and the first one for the coastal part of this region.

Order Perciformes, Family Acanthuridae

Acanthurus coeruleus Bloch & Schneider, 1801 (Figure 7E). One juvenile individual was recorded at the approximate depth of 8 m, Deserta Island, Arvoredo Marine Biological Reserve, Santa Catarina in 2010, and one adult (ca. 30 cm total length) was recorded at a depth of 12 m at Saco do Farol, Arvoredo Island in

March 2014. *Remarks:* Previous southernmost record for this species wasin the state of São Paulo (Carvalho-Filho 1999; Moura et al. 1999).

Acanthurus monroviae Steindachner, 1876. One individual, an adult male, was recorded at the approximate depth of 7 m at Parcel da Deserta, Arvoredo Marine Biological Reserve, Santa Catarina in February 2015. *Remarks:* This individual represents the second record for the Western Atlantic Ocean, being previously recorded for Laje de Santos, in the state of São Paulo (Luiz et al. 2004, 2010). Its native range is in the Eastern Atlantic.

Family Apogonidae

Apogon americanus (Castelnau 1855) (Figure 7F). One individual was recorded at a depth of 10 m at Saco do Farol, Arvoredo Island in February 2014. *Remarks:* Previous southernmost record was in the state of São Paulo (Carvalho-Filho 1999).

Family Carangidae

Decapterus punctatus (Cuvier, 1829). Many individuals were seen and photographed at 10–15 m deep, at Deserta Island, Santa Catarina, 2015. *Remarks:* The previous southernmost record was in the state of São Paulo (Carvalho-Filho 1999).

Family Chaetodontidae

Chaetodon sedentarius Poey, 1860 (Figure 8A). One individual was recorded at the approximate depth of 9 m, Arvoredo Marine Biological Reserve, Santa Catarina in 2010. *Remarks:* Previous southernmost record was in the state of São Paulo (Carvalho-Filho 1999).

Family Labridae

Clepticus brasiliensis Heiser, Moura & Robertson, 2000 (Figure 8B). One individual was recorded at the approximate depth of 6 m, Deserta Island, Arvoredo Marine Biological Reserve, Santa Catarina, in April 2011. *Remarks:* The previous southernmost record was in the state of São Paulo (Carvalho-Filho 1999).

Family Pomacentridae

Chromis flavicauda (Günther, 1880)(Figure 8E). Three individuals were recorded at the approximate depth of 10 m, Xavier Island, Santa Catarina in February 2011. One of these individuals was resighted in April of the same year. *Remarks:* Previous southernmost record was in the state of São Paulo (Carvalho-Filho 1999).

Stegastes partitus (Poey, 1868) (Figure 8C and 8D). One individual was recorded at the approximate depth

of 6 m, Galé Island, Arvoredo Marine Biological Reserve, Santa Catarina in February 2013. *Remarks:* The individual recorded represents the first record for the Southern Atlantic Ocean. Its native range is in the Northwestern Atlantic, south to Venezuela (Cervigón 1993).

Order Tetraodontiformes, Family Monacanthidae

Cantherhines macrocerus (Hollard, 1853) (Figure 8F). One individualrecorded being cleaned by a juvenile of *Pomacanthus paru* at the approximate depth of 2 m at

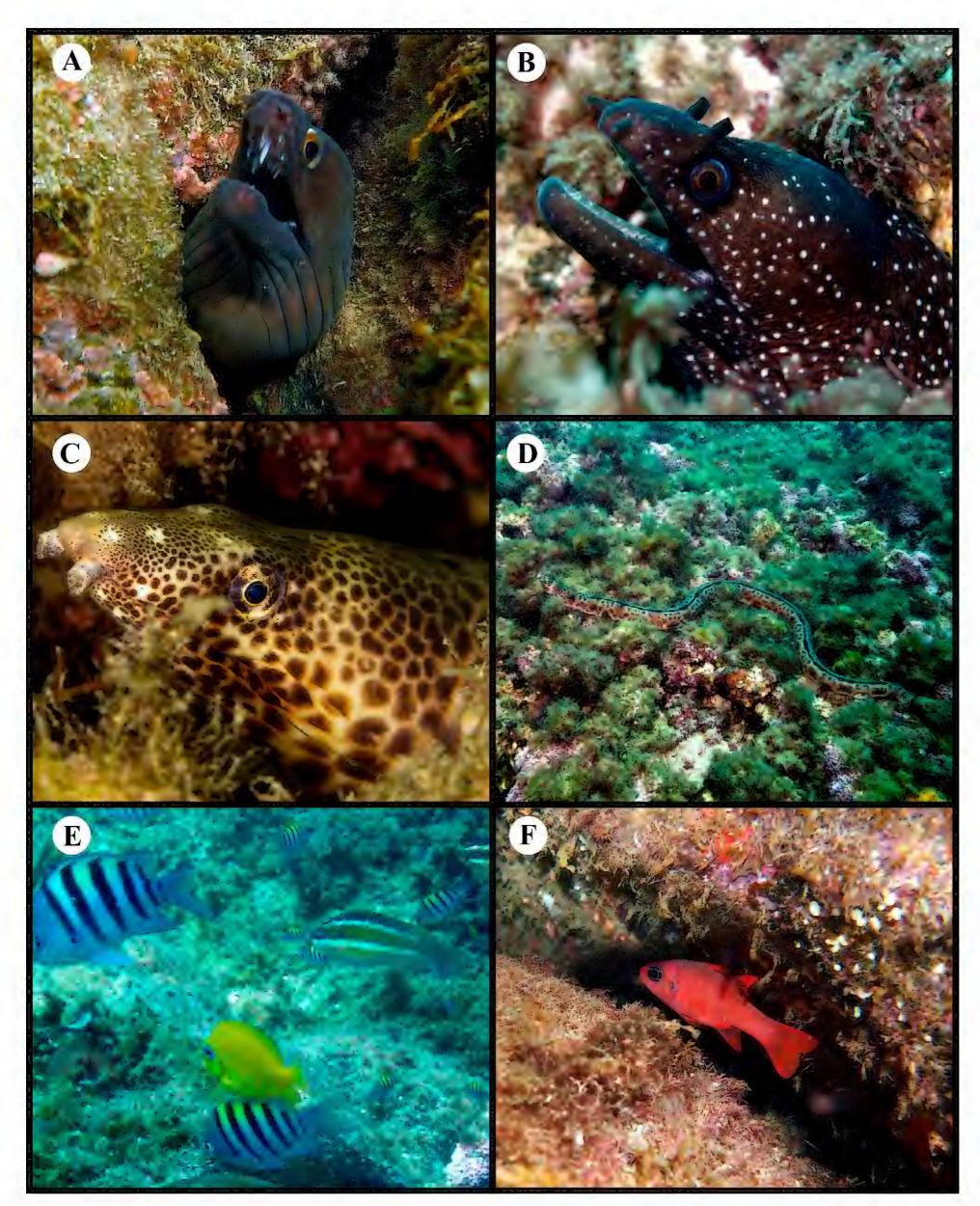


Figure 7. Examples of new records of reef fish species to the state of Santa Catarina, Southern Brazil. (**A**) Purplemouth Moray, *Gymnothorax vicinus*; (**B**) Reticulate Moray, *Muraena retifera*; (**C–D**) Ornate Snake Eel, *Herpetoichthys regius*; (**E**) Blue Tang Surgeonfish, *Acanthurus coeruleus*; (**F**) Brazilian Flamefish, *Apogon americanus*.

Praia da Sepultura, Bombinhas, Santa Catarina in March 2015. *Remarks:* Previous southernmost record was in the state of São Paulo (Carvalho-Filho 1999).

A growing number of reef fishes have been detected outside of their native range in the Atlantic (Freitas et al. 2013; Luiz et al. 2013), with at least three recent examples reaching south-southeastern Brazil: the Azores Chromis, *Chromis limbata* (Valenciennes, 1833), the West African Tang, *Acanthurus monroviae* Steindachner, 1876, and the Bannerfish, *Heniochus acuminatus* (Linnaeus, 1758). The

first two species are known from the Eastern Atlantic and have reached the Brazilian coast after breaching the Mid-Atlantic Barrier (Luiz et al. 2004; Leite et al. 2009). The Bannerfish is distributed all over the Indo-Pacific and it is controversial whether its arrival in Brazil means a long dispersal via South Africa or an aquarium release (Luiz et al. 2014). From these three species, only *C. limbata* has successfully established populations in the Southwestern Atlantic (Anderson *et al.* pers. obs.). Of the new records presented by this work, the Bicolor Damselfish, *Stegastes partitus*, and the Ornate Snake



Figure 8. Examples of new records of reef fish species to the state of Santa Catarina, Southern Brazil. (**A**) Reef Butterflyfish, *Chaetodon sedentarius*; (**B**) Brazilian Creolewrasse, *Clepticus brasiliensis*; (**C–D**) Bicolor Damselfish, *Stegastes partitus*; (**E**) Cobalt Chromis, *Chromis flavicauda*; (**F**) Whitespotted Filefish, *Cantherhines macrocerus*.

Eel, Herpetoichthys regius, are considerably unnusual not only because they expand species' known geographic range by several thousands of kilometers, but also because of large environmental differences between their place of origin and the Santa Catarina coast.

The Bicolor Damselfish is a Northwestern Atlantic endemic species, ranging from Florida to Venezuela (Humannn and Deloach 2014). This species is heavily associated with coral reefs, even when occurs in cooler upwelling Caribbean locations, such as the Venezuelan coast, and therefore its occurrence on a transitional zone with no coral reef builders (see Castro and Pires 2001, for distribution of coral reefs in Brazil) is at least intriguing. Introduction via ship's ballast water is unlikely because most fish larvae do not survive for long periods in that environment (Carlton 1985). Aquarium trade is also unlikely because this species is not commonly exported from the Caribbean, and there are no records of it being commercialized in the study region. The building of new platforms and intense oil industry activity seems to be the more likely form of artificial introduction because movement of these structures is intensifying worldwide and adults from some species are known to have been transported through those means (Dulcic and Dragicevic 2013). However, the possibility of a natural colonization cannot be dismissed and only future monitoring of this species presence along the Brazilian coast could clarify this range extension causes.

The Ornate Snake Eel is considered to be endemic to the Eastern Atlantic and islands of Santa Helena, Ascension and Saint Paul's Rocks (Wirtz et al. 2015), none of which is located less than 3,700 km from Santa Catarina coast. Although no ecological information concerning this species was found in the literature, in the two occasions it was encountered in Santa Catarina, it wandered through shallow rhodolith beds. Contrary to Stegastes partitus, it is highly unlikely that this species was artificially introduced at Santa Catarina coast and, so, natural dispersal might account for its presence there. The first and most likely hypothesis is that it is also present in other places along the Brazilian coast, but was overlooked due to its cryptic nature and, especially, to its poorly sampled habitat (rhodolith beds), for which Santa Catarina is the southernmost limit (Gherardi 2004; Pascelli et al. 2013). The alternative hypothesis is that it represents another case of extreme dispersal event driven by Brazil Current.

Zoogeography, tropical affinities and remarkable absences

Most of the recorded species (162 species or 58.3%) are known to occur only in the Western Atlantic Ocean. Species occurring on both sides of the Atlantic Ocean (Western and Eastern Atlantic, 60 species) total 21.6% of the richness. Circumtropical cosmopolitan species (24 species) totaling 8.6% and 15 species were considered

circumglobal cosmopolitans species (5.4%) (Floeter et al. 2008; Froese and Pauly 2014).

Also, 23 of the recorded species (8.3%) are endemic to the Brazilian Province (Figure 9), a little lower than the number for the whole Brazilian coast (10.5%; Floeter et al. 2008). This might be due to the fact that many endemic Brazilian species possess tropical affinities, with some attaining their southern limit of occurrence northwards, in the states of Rio de Janeiro or São Paulo (Carvalho-Filho 1999; Luiz et al. 2009). In fact, all these localities are part of the Southwestern Atlantic Shelf (Matano et al. 2010) and are affected by seasonal cool waters that can restrict the occurrence of tropical species. Examples of endemic tropical species that do not reach Santa Catarina are *Haemulon squamipinna*, *Halichoeres penrosei*, and *Lutjanus alexandrei*, just to mention a few.

Despite some tropical reef fish species that do not occur southward to Santa Catarina state, many do and overall its faunal domain can be considered tropical with the enrichment of temperate elements. These temperate elements are, however, less important for species compostion than its relatively high latitude would predict. This is particularly true when comparing to southeastern region reefs that are heavily affected by upwelling, such as Arraial do Cabo and Ilha Rasa in Rio de Janeiro state, and Laje de Santos in São Paulo state (Luiz et al. 2008; Carvalho-Filho et al. 2009; Bertonciniet al. 2013). In these places, temperate species that occur in deep waters throughout the Brazilian shelf have been recorded for shallow waters associated with frequent upwelling events.

This upwelling of deep water/temperate species includes the occurrence of *Dules auriga* Cuvier, 1829, Pagrus pagrus (Linnaeus, 1758), Pinguipes brasilianus Cuvier, 1829, Halichoeres sazimai Luiz, Ferreira & Rocha, 2009, Acanthistius brasilianus (Cuvier, 1828) and Pronotogrammus martinicensis (Guichenot, 1868) for depths shallower than 40 m (Irigoyen et al. 2008; Luiz et al. 2008; Carvalho-Filho et al. 2009; Bertonciniet al. 2013). The first three species are common in shallow Argentine reefs (Irigoyen and Galván 2010), H. sazimai have its southern distribution limit at Santa Catarina (Barneche et al. 2009), but *P. martinicensis* have its only known shallow water population in these upwelling areas along the "Arc of Capricorn" (Carvalho-Filho et al. 2009) and A. brasilianus in this region and also Uruguay (Irigoyen et al. 2008; Irigoyen et al. 2010). Dules auriga is commonly found in Santa Catarina southward from Xavier Island (27°36′ S, 048°23′ W), while H. sazimai and Pagrus pagrus are rare or uncommon, even in shallow habitats in the southern part of the state (RM pers obs). But the most intriguing issue is that Pinguipes brasilianus, Acanthistius brasilianus and Pronotogrammus martinicensis have never been recorded for the shallow reefs despite relatively wide geographic sampling and the fact that the higher latitude reefs of Santa Catarina are exposed to temperatures sometimes as low as these upwelling places northward. Recently, *Pronotogrammus martinicensis* was recorded from Santa Catarina at a depth of 130 m, associated with the sunken german

submarine U-513 (Figure 10). This suggests that other temperate species might also be present in deep reefs, and may be confirmed by more observations and further sampling of these habitats.

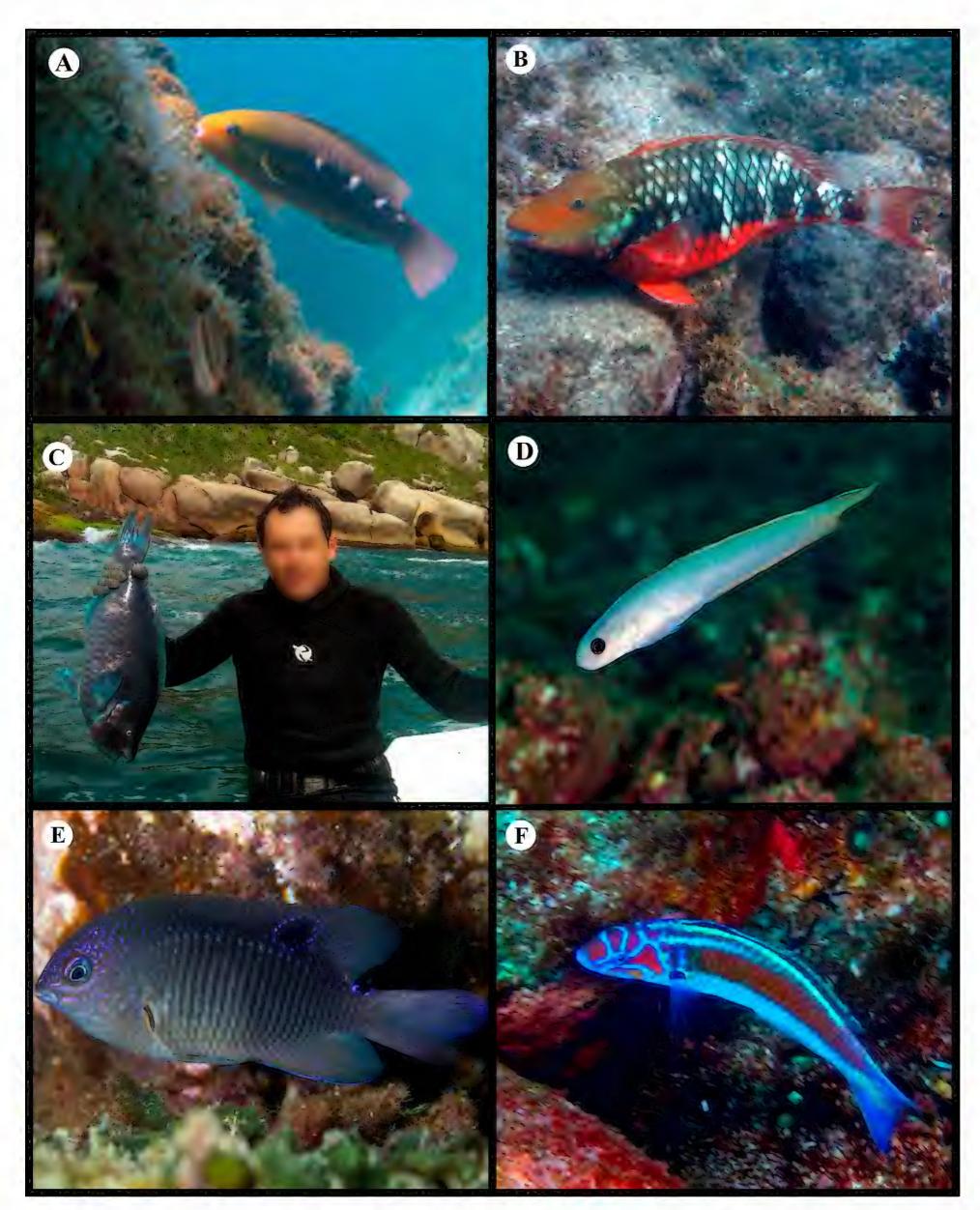


Figure 9. Examples of endemic Brazilian reef fish species observed in reefs of Santa Catarina, Southern Brazil. (A) Striped Parrotfish, *Scarus zelindae*; (B) Reef Parrotfish, *Sparisoma amplum*; (C) Greenback Parrotfish, *Scarus trispinosus*; (D) Brazilian Dartfish, *Ptereleotris randalli*; (E) Brazilian Dusky Damselfishes, *Stegastes fuscus*; (F) Noronha Wrasse, *Thalassoma noronhanum*.



Figure 10. Pronotogrammus martinicensis recorded at 130 m depth associated with the sunken German submarine U-513 off Santa Catarina, Southern Brazil.

Conclusions

The ichthyofauna of Santa Catarina state is mostly characterized by tropical species brought from the northern warm waters by oceanic currents. Nevertheless, the cold waters in the austral winter sea surface temperature can affect the survival of such species, leading to the extirpation of populations (cf. Bohnsack 1983; Hsieh et al. 2008). The unusual new records presented in the present work indicates that long-term monitoring studies will allow a better understanding of connectivity patterns along the coast of Santa Catarina as well as the possible establishment of new populations at their southernmost limit of distribution. Of interest also are cold-water related species whose reasons for the disjunct observed distributions, which exclude Santa Catarina, are not clear. Further sampling efforts on deeper habitats might reveal novel insights concerning their distribution status.

ACKNOWLEDGEMENTS

We thank John E. McCosker for helping in the identification of *Herpetoichthys regius* (Richardson, 1848). Photographs: E. Bastos for *H. regius*, E. Faria-Júnior for *Apogon americanus*, A.F. Sarti for *Genidens genidens*, A.M.R. Liedke for *Centropyge aurantonotus*, R.M. Bonaldo for *Ptereleotris randalli* and A. Dutra for *Thalassoma noronhanum*. Cristian Dimitrius kindly provided photographs of *Pronotogrammus martinicensis* from the U-513 submarine. We would also like to thank the colleagues that contributed information, discussions, references or participated during surveys: D.R. Barneche, E. Bastos, D.F. Dinslaken, G.O. Longo, E. Faria-Júnior, A.G.V. Floeter, L. Fontoura, J.P. Krajewski, G.C. Ribeiro and F.F. Pacheco for old literature on Santa Catarina reef fishes.Funding sources: SISBIOTA-Mar (PI: S.R.F., CNPq

563276/2010-0; FAPESC 6308/2011-8), Projeto Ilhas do Sul (PI: S.R.F., CNPq 475367/2006-5), Projeto MAArE – Monitoramento Ambiental do Arvoredo e Entorno (PI: Barbara Segal) is a condition set by the ICMBio in the context of IBAMA's environmental licensing process), CAPES (A.B.A., R.A.M., L.T.N., J.P.Q.) and Projeto Biodiversidade Marinha do Estado de Santa Catarina (PI: Alberto Lindner, FAPESC 4302/2010-8).

LITERATURE CITED

Acha, E.M., H.W. Mianzan, R.A. Guerrero, M. Favero and J. Bava. 2004. Marine fronts at the continental shelves of austral South America: physical and ecological processes. Journal of Marine Systems 44: 83–105. doi: 10.1016/j.jmarsys.2003.09.005

Anderson, A.B., L. Fontoura, G.O. Longo and S.R. Floeter. 2014a. Peixes; pp. 70–89, in: A. Lindner (ed.). Vida Marinha de Santa Catarina. Florianópolis: Editora da Universidade Federal de Santa Catarina.

Anderson, A.B., R.M. Bonaldo, D.R. Barneche, C.W. Hackradt, F.C. Félix-Hackradt, J.A. García-Chartón and S.R. Floeter. 2014b. Recovery of grouper assemblages indicates effectiveness of a marine protected area in Southern Brazil. Marine Ecology Progress Series 514: 207–215. doi: 10.3354/meps11032

Barneche, D.R., A.B. Anderson, S.R. Floeter, M. Silveira, D.F. Dinslaken and A. Carvalho-Filho. 2009. Ten new records of reef fish on the coast of Santa Catarina State, Brazil. Marine Biodiversity Records 2(143): 1–4. doi: 10.1017/S1755267209990613

Bernardi, G., R. Noguchi, A.B. Anderson, S.R. Floeter and C.E.L. Ferreira. 2013. Sargo Amarelo, a traditionally recognized hybrid between two species of Brazilian reef fishes. Marine Biodiversity 43(4): 255–256. doi: 10.1007/s12526-013-0169-0

Bertoncini, A., C. Rangel, L. Chaves, J. Mendonça-Neto and C. Monteiro-Neto. 2013. Peixes recifais do monumento natural das Ilhas Cagarras; pp. 106–137, in: Morais, F., A. Bertocini and A. Aguiar (eds.). História, Pesquisa e Biodiversidade do Monumento Natural das Ilhas Cagarras. Rio de Janeiro: Museu Nacional.

Boehm, J.T., L. Woodall, P.R. Teske, S.A. Lourie, C. Baldwin, J. Waldman and M. Hickerson. 2013. Marine dispersal and barriers drive Atlantic seahorse diversification. Journal of Biogeography 40(10): 1839–1849. doi: 10.1111/jbi.12127

- Bohnsack, J.A. 1983. Resiliency of reef fish communities in the Florida Keys following a January 1977 hypothermal fish kill. Environmental Biology of Fishes 9(1): 41–53. doi: 10.1007/BF00001057
- Boschi, E.E. 2000. Species of decapod crustaceans and their distribution in the American marine zoogeographic provinces. Revista de Investigacion y Desarrollo Pesquero 13: 7–64. doi: http://www.oceandocs.org/handle/1834/2606
- Briggs, J.C. and B.W. Bowen. 2012. A realignment of marine biogeographic provinces with particular reference to fish distributions. Journal of Biogeography 39: 12–30. doi: 10.1111/j.1365-2699.2011.02613.x
- Capel, K.C.C., B. Segal, P. Bertuol and A. Lindner. 2012. Corallith beds at the edge of the tropical South Atlantic. Coral Reefs 31: 75. doi: 10.1007/s00338-011-0818-3
- Carlton, J.T. 1985. Transoceanic and interoceanic dispersal of coastal marine organisms: the biology of ballast water. Oceanography and Marine Biology: an Annual Review 23: 313–371. doi: http://research.mblwhoilibrary.org/works/7187
- Carvalho, J.L.B., C.A.F. Schettini and T.M. Ribas. 1998. Estrutura termohalina do litoral centro-norte catarinense. Notas Técnicas da Faculdade de Ciências do Mar 2: 181–197. doi: 10.14210/bjast. v2n1.p181-197
- Carvalho-Filho, A. 1999. Peixes: Costa Brasileira. São Paulo: Editora Melro. 320 pp.
- Carvalho-Filho, A., C.E.L. Ferreira and M. Craig. 2009. A shallow water population of *Pronotogrammus martinicensis* (Guichenot, 1868) (Teleostei: Serranidae: Anthiinae) from South-western Atlantic, Brazil. Zootaxa 2228: 29–42. doi: http://www.mapress.com/zootaxa/2009/f/zo2228po42f.pdf
- Castro, C.B., D.O. Pires. 2001. Brazilian coral reefs: what we already know and what is still missing. Bulletin of Marine Science 69(2): 357–371. http://www.ingentaconnect.com/content/umrsmas/bullmar/2001/0000069/00000002/art00013
- Cervigón, F. 1993. Los Peces Marinos de Venezuela. Vol. 2. Caracas: Fundación Científica Los Roques. 497 pp.
- Charrid, R.Jr. 2011. The holoplankton of the Santa Catarina coast, Southern Brazil. Annals of the Brazilian Academy of Sciences 83(2): 575–588. doi: 10.1590/S0001-37652011000200017
- Choat, J.H., O.S. Klanten, L. Van Herwerden, D.R. Robertson and K.D. Clements. 2012. Patterns and processes in the evolutionary history of parrotfishes (family Labridae). Biological Journal of the Linnean Society 107(3): 529–557. doi: 10.1111/j.1095-8312.2012.01959.x
- Craig, M.T. and P.A. Hastings. 2007. A molecular phylogeny of the groupers of the subfamily Epinephelinae (Serranidae) with a revised classification of Epinephelini. Ichthyological Research 54(1): 1–17. doi: 10.1007/s10228-006-0367-x
- Craig, M.T., Y.J.S. Mitcheson and P.C. Heemstra. 2011. Groupers of the world: a field and market guide. Grahamstown: CRC Press. 403 pp.
- Diehl, F.L. and N.O. Horn Filho. 1996. Compartimentação geológicogeomorfológica da zona litorânea e planície costeira do Estado de Santa Catarina. Notas Técnicas da Faculdade de Ciências do Mar 9: 39–50.
- Dulcic, J. and B. Dragicevic. 2013. *Holacanthus ciliaris* (Linnaeus, 1758) (Teleostei: Pomacanthidae), first record from the Mediterranean Sea. Journal of Applied Ichthyology 29(2): 465–467. doi: 10.1111/jai.12096
- Edwards, A.J. and R. Lubbock. 1983. Marine zoogeography of Saint Paul's Rocks. Journal of Biogeography 10(1): 65–72. http://www.jstor.org/stable/2844583
- Faircloth, B.C., L. Sorenson, F. Santini and M.E. Alfaro. 2013. A phylogenomic perspective on the radiation of ray-finned fishes based upon targeted sequencing of ultraconserved elements (UCEs). PLOS One 8(6): e65923. doi: 10.1371/journal. pone.0065923

- Feitoza, B.M., L.A. Rocha, O.J. Luiz, S.R. Floeter and J.L. Gasparini. 2003. Reef fishes of St. Paul's Rocks: new records and notes in biology and zoogeography. International Journal of Ichthyology and Aquatic Biology 7(2): 61–82.
- Ferreira, C.E.L., S.R. Floeter, J.L. Gasparini, B.P. Ferreira and J.-C. Joyeux. 2004. Trophic structure patterns of Brazilian reef fishes: a latitudinal comparison. Journal of Biogeography 31(7): 1093–1106. doi: 10.1111/j.1365-2699.2004.01044.x
- Floeter, S.R., R.Z.P. Guimarães, L.A. Rocha, C.E.L. Ferreira, C.A. Rangel and J.L. Gasparini. 2001. Geographic variation in reef-fish assemblages along the Brazilian coast. Global Ecology and Biogeography 10(4): 423–433. doi: 10.1046/j.1466-822X.2001.00245.x
- Floeter, S.R., L.A. Rocha, D.R. Robertson, J.C. Joyeux, W.F. Smith-Vaniz, P. Wirtz, A.J. Edwards, J.P. Barreiros, C.E.L. Ferreira, J.L. Gasparini, A. Brito, J.M. Falcón, B.W. Bowen and G. Bernardi. 2008. Atlantic reef fish biogeography and evolution. Journal of Biogeography 35(1): 22–47. doi: 10.1111/j.1365-2699.2007.01790.x
- Frable, B.W., C.C. Baldwin, B.M. Luther and L.A. Weigt. 2013. A new species of Western Atlantic lizardfi fish (Teleostei: Synodontidae: *Synodus*) and resurrection of *Synodus bondi* Fowler, 1939, as a valid species from the Caribbean with redescriptions of *S. bondi*, *S. foetens* (Linnaeus, 1766), and *S. intermedius* (Agassiz, 1829). Fishery Bulletin 111: 122–146. doi: 10.7755/FB.111.2.2
- Freitas, R., O.J. Luiz, P.N. Silva, S.R. Floeter, G. Bernardi and C.E.L. Ferreira. 2014. The occurrence of *Sparisoma frondosum* (Teleostei: Labridae) in the Cape Verde Archipelago, with a summary of expatriated Brazilian endemic reef fishes. Marine Biodiversity 44(2): 173–179. doi: 10.1007/s12526-013-0194-z
- Froese, R. and D. Pauly. 2014. FishBase. Version 2014. Accessed at http://www.fishbase.org, 22 March 2014.
- Galván, D.E., L.A. Venerus and A.J. Irigoyen. 2009. The Reef-fish Fauna from the Northern Patagonian Gulfs of Argentina, Southwestern Atlantic. The Open Fish Science Journal 2(1): 90–98. doi: 10.2174/1874401X00902010090
- Gherardi, D.F.M. 2004. Community structure and carbonate production of a temperate rhodolith bank from Arvoredo Island, Southern Brazil. Brazilian Journal of Oceanography 52(3/4): 207–224. doi: 10.1590/S1679-87592004000300004
- Godoy, M.P. 1987. Peixes do estado de Santa Catarina. Florianópolis: Editora da Universidade Federal de Santa Catarina. 572 pp.
- Greenacre, M.J. 2007. Correspondence analysis in practice, 2nd edition. Boca Raton: Chapman & Hall/CRC Press. 296 pp.
- Hackradt, C.W. and F.C. Félix-Hackradt. 2009. Assembleia de peixes associados a ambientes consolidados no litoral do Paraná, Brasil: uma análise qualitativa com notas sobre sua bioecologia. Papéis Avulsos de Zoologia 49(31): 389–403. doi: 10.1590/S0031-10492009003100001
- Hille, E., C.A.F. Schettini and M.R. Ribeiro. 2008. Estrutura termohalina no litoral de Santa Catarina nos anos de 2005 e 2006; pp. 371–381, in: E.S. Braga (ed.). Oceanografia e mudanças globais. São Paulo: Editora da Universidade de São Paulo.
- Hostim-Silva, M., A.B. Andrade, L.F. Machado, L.C. Gerhardinger, F.A. Daros, J.P. Barreiros and E.A.S. Godoy. 2006. Peixes de Costão Rochoso de Santa Catarina: Ilha do Arvoredo. Itajaí: Editora da Universidade do Vale do Itajaí. 134 pp.
- Hsieh, C.-J., K.-W. Chang, C.-J. Lin, S.S. Keerthi and S. Sundararajan. 2008. A dual coordinate descent method for large-scale linear SVM. Proceedings of the 25th International Conference on Machine Learning: 408–415. doi: 10.1145/1390156.1390208
- Humann, P. and N. DeLoach. 2014. Reef fish identification: Florida, Caribbean, Bahamas, 4th edition. Jacksonville, Florida: New World Publications. 537 pp.
- Irigoyen, A.J., Gerhardinger L.C. and A. Carvalho-Filho. 2008. On the status of the species of *Acanthistius* (Gill, 1862) (Percoidei)

- in the South-West Atlantic Ocean. Zootaxa 1813: 51–59. http://www.mapress.com/zootaxa/2008/2/zto1813p059.pdf
- Irigoyen, A.J. and D.E. Galván. 2010. Peces de arrecife argentinos. Puerto Madryn: Proyecto Arrecife. 90 pp.
- Irigoyen, A.J., Y. Marin and A. Carvalho-Filho. 2010. Occurrence of *Acanthistius brasilianus* (Cuvier, 1828) in Uruguayan waters (35°45′S): when poor taxonomy means poor ecological knowledge. Journal of Applied Ichthyology, 26: 600–601. doi: 10.1111/j.1439-0426.2010.01390.x
- IUCN (International Union for Conservation of Nature). 2013. Red List of threatened species. Accessed at http://www.iucnredlist.org, 15 January 2015.
- Knudsen, S.W. and K.D. Clements. 2013. Revision of the family Kyphosidae (Teleostei: Perciformes). Zootaxa 3751(1): 001–101. doi: 10.11646/zootaxa.3751.1.1
- Leite, J.R., A.A. Bertoncini, L. Bueno, F. Daros, J. Alves and M. Hostim-Silva. 2009. The occurrence of Azores Chromis, *Chromis limbata* in the South-western Atlantic. Marine Biodiversity Records 2: e145. doi: 10.1017/S1755267209990637
- Lema, T. 1976. Ocorrência de várias espécies de peixes tropicais marinhos na costa do Estado de Santa Catarina, Brasil (Osteichthyes, Actinopterygii, Teleostei). Iheringia Série Zoologia 49: 39–65. http://www.biodiversitylibrary.org/item/106278
- Lema, T., C.A.S. Lucena and Z.M.S. Lucena. 1980. Novas adendas à ictiofauna marinha do extremo sul do Brasil (Actinopterygii: Teleostei). Iheringia Série Zoologia 56: 103–120. http://www.biodiversitylibrary.org/item/107517
- Luiz, O.J., S.R. Floeter, J.L. Gasparini, C.E.L. Ferreira and P. Wirtz. 2004. The occurrence of *Acanthurus monroviae* (Perciformes:Acanthuridae) in the South-Western Atlantic, with comments on other Eastern Atlantic reef fishes occurring in Brazil. Journal of Fish Biology 65: 1173–1179. doi: 10.1111/j.1095-8649.2004.00519.x
- Luiz, O.J., A. Carvalho-Filho, C.E.L. Ferreira, S.R. Floeter, J.L. Gasparini and I. Sazima. 2008. The reef fish assemblage of the Laje de Santos Marine State Park, Southwestern Atlantic: annotated checklist with comments on abundance, distribution, trophic structure, symbiotic associations, and conservation. Zootaxa 1807: 1–25. http://www.mapress.com/zootaxa/2008/f/zo1807p025f.pdf
- Luiz, O.J., C.E.L. Ferreira and L.A. Rocha. 2009. *Halichoeres sazimai*, a new species of wrasse (Perciformes: Labridae) from the Western South Atlantic. Zootaxa 2092: 37–46. http://www.mapress.com/zootaxa/2009/f/zo2092p046f.pdf
- Luiz, O.J., I. Sazima, L.F. Waib and C.E.L. Ferreira. 2010. A honeymoon in Brazil: the spawning behavior of an exotic reef fish in the Western South Atlantic. Neotropical Ichthyology 8:369–371. http://www.scielo.br/pdf/ni/v8n2/v8n2a16.pdf
- Luiz, O.J., S.R. Floeter, L.A. Rocha and C.E.L. Ferreira. 2013. Perspectives for the lionfish invasion in the South Atlantic: Are Brazilian reefs protected by the currents? Marine Ecology Progress Series 485: 1–7. doi: 10.3354/meps10383
- Luiz, O.J., E. Comin and J.S. Madin. 2014. Far away from home: the occurrence of the Indo-Pacific Bannerfish *Heniochus acuminatus* in the Atlantic. Bulletin of Marine Science 90(2): 741–744. doi: 10.5343/bms.2013.1046
- Matano, R.P., E.D. Palma and A.R.Piola. 2010. The influence of the Brazil and Malvinas currents on the Southwestern Atlantic shelf circulation. Ocean Science Discussions 7: 837–871. doi: 10.5194/0s-6-983-2010
- Molina-Schiller, D., S.A. Rosales and T.R.O. Freitas. 2005. Oceanographic conditions off coastal South America in relation to the distribution of Burmeister's porpoise, (*Phocoena spinipinnis*). Latin American Journal of Aquatic Mammals 4(2): 141–156. doi: 10.5597/lajamooo78

- Möller Jr, O.O., A.R. Piola, A.C. Freitas and E.J.D. Campos. 2008. The effects of river discharge and seasonal winds on the shelf off Southeastern South America. Continental Shelf Research 28(13): 1607–1624. doi: 10.1016/j.csr.2008.03.012
- Moura, R.L., J.L. Gasparini and I. Sazima. 1999. New records and range extensions of reef fishes in theWestern South Atlantic, with comments on reef fish distribution along the Brazilian coast. Revista Brasileira de Zoologia 16(2): 513–530. doi: 10.1590/S0101-81751999000200017
- Near, T.J., R.I. Eytan, A. Dornburg, K.L. Kuhn, J.A. Moore, M.P. Davis, P.C. Wainwright, M. Friedman and W.L. Smith. 2012. Resolution of ray-finned fish phylogeny and timing of diversification. Proceedings of the National Academy of Sciences of the United States of America, 109(34): 13698–13703. doi: 10.1073/pnas.1206625109
- Nenadic, O. and M. Greenacre. 2007. Correspondence analysis in R, with two and three-dimensional graphics: The ca package. Journal of Statistical Software 20(3): 1–13. http://www.jstatsoft.org/v2o/io3/paper/
- Pascelli, C., P. Riul, R. Riosmena-Rodríguez, F. Scherner, M. Nunes, J.M. Hall-Spencer, E.C. de Oliveira and P. Horta. 2013. Seasonal and depth-driven changes in rhodolith bed structure and associated macroalgae off Arvoredo island (southeasternBrazil). Aquatic Botany 111: 62–65. doi: 10.1016/j.aquabot.2013.05.009
- Piola, A.R., E.J.D. Campos, O.O. Möller Jr., M. Charo and C. Martinez. 2000. Subtropical shelf front of Eastern South America. Journal of Geophysical Research 105(C3): 6565–6578. doi: 10.1029/1999JC000300
- Piola, A.R., R.P. Matano, E.D. Palma, O.O. Möller Jr. and E.J.D. Campos. 2005. The influence of the Plata River discharge on the Western South Atlantic shelf. Geophysical Research Letters 32(1): 1–4. doi: 10.1029/2004GL021638
- Pita, P., D. Fernández-Márquez and J. Freire. 2014. Short-term performance of three underwater sampling techniques for assessing differences in the absolute abundances and in the inventories of the coastal fish communities of the Northeast Atlantic Ocean. Marine and Freshwater Research 65(2): 105–113. doi: 10.1071/MF12301
- Randall, J.E. 1967. Food habits of reef fishes of the West Indies. Studies in Tropical Oceanography5: 665–847. http://www.aoml.noaa.gov/general/lib/CREWS/Cleo/PuertoRico/prpdfs/randall-habits.pdf
- Randall, J.E. 1996. Caribbean Reef Fishes. Neptune City: TFH Publications. 368 pp.
- Rosa, R.S. and R.L. Moura. 1997. Visual assessment of reef fish community structure in the Atol das Rocas Biological Reserve, off northeastern Brazil. Proceedings of the 8th International Coral Reef Symposium 1: 983–986.
- Sazima, I. 2006. Similarities in feeding behaviour between some marine and freshwater fishes in two tropical communities. Journal of Fish Biology 29: 53–65. doi: 10.1111/j.1095-8649.1986. tb04926.x
- Seeliger, U., C. Odebrecht and J.P. Castello. 1997. Subtropical convergence environments: the coast and sea in the southwestern Atlantic. Berlim: Springer. 308 pp.
- Silveira, R.B., R. Siccha-Ramirez, J.R.S. Silva and C. Oliveira. 2014. Morphological and molecular evidence for the occurrence of three *Hippocampus* species (Teleostei: Syngnathidae) in Brazil. Zootaxa 3861: 317–332. doi: 10.11646/zootaxa.3861.4.2
- Simon, T., R.M. Macieira and J.-C. Joyeux. 2013. The shore fishes of the Trindade–Martin Vaz insular complex: an update. Journal of Fish Biology 82(6): 2113–2127. doi: 10.1111/jfb.12126
- Smith, W.L. and M.T. Craig. 2007. Casting the Percomorph net widely: the importance of broad taxonomic sampling in the search for the placement of the serranid and percid fishes. Copeia 1: 35–55. 10.1643/0045-8511(2007)7[35:CTPNWT]2.0.CO;2

Sobrinho, R.J. de S., A. Bresolin and R.M. Klein. 1969. Os manguezais na ilha de Santa Catarina. Insula 2: 1–21.

Spalding, M.D., H.E. Fox, G.R. Allen, N. Davidson, Z.A. Ferdaña, M. Finlayson, B.S. Halpern, M.A. Jorge, A. Lombana, S.A. Lourie, K.D. Martin, E. McManus, J. Molnar, C.A. Recchia and J. Robertson. 2007. Marine ecoregions of the world: a bioregionalization of coastal and shelf areas. BioScience 57(7): 573–583. doi: 10.1641/B570707

Westneat, M.W. and M.E. Alfaro. 2005. Phylogenetic relationships and evolutionary history of the reef fish family Labridae. Molecular Phylogenetics and Evolution 36(2): 370–390. doi: 10.1016/j.ympev.2005.02.001

Wirtz, P., J. Bingeman, J. Bingeman, R. Fricke, T. Hook and J. Young.

2015. The fishes of Ascension Island, central Atlantic Ocean – new records and an annotated check-list. Journal of the Marine Biological Association of the United Kingdom [advance access]. doi: 10.1017/S0025315414001301

Authors' contribution statement: All authors conceived the study, collected field, museum and literature data, analyzed the data and wrote the paper.

Received: 19 February 2015 **Accepted:** 28 May 2015

Academic editor: Osmar J. Luiz

Appendix 1

Table A1. Vouchers specimens from MZUSP (Museu de Zoologia da Universidade de São Paulo, Brazil); vouchers from NEMAR (Center of Sea Studies of Universidade Federal de Santa Catarina UFSC / Brazil); vouchers from CIUFSC (Ichthyological collection of Universidade Federal de Santa Catarina, UFSC / Brazil); voucher from USNM (National Museum of Natural History; Smithsonian Institution; Washington, DC) and voucher from UFRGS (Universidade Federal do Rio Grande do Sul). Names in parenthesis are the previous recognized as valid for the area.

Family	Species	MZUSP	CIUFSC	NEMAR
Acanthuridae	Acanthurus bahianus	MZUSP55397	CUFSC1391	-
Acanthuridae	Acanthurus chirurgus	MZUSP55341	-	2
Ariidae	Cathorops spixii	-	CIUFSC251	ARII 1982.001.191
Ariidae	Genidens barbus	-	CIUFSC1079	ARII 1992.029.219
Ariidae	Genidens genidens	-	CIUFSC1329	ARII 1982.002.192
Balistidae	Balistes capriscus	-	CIUFSC814	BALIS 1981.002.1186
Batrachoididae	Porichthys porosissimus	-	CIUFSC779	BATRA 1982.001.101
Belonidae	Strongylura marina	-	-	BELO 1992.011.138
Blenniidae	Hypleurochilus fissicornis	MZUSP55318	-	BLEN 1984.002.121
Blenniidae	Hypleurochilus pseudoequipinnis	MZUSP55320	-	-
Blenniidae	Hypsoblennius invemar	MZUSP55319	-	_
Blenniidae	Ophioblennius trinitatis	MZUSP55452	_	-
Blenniidae	Parablennius marmoreus	MZUSP55451	-	-
Blenniidae	Parablennius pilicornis	-	CIUFSC605	BLEN 1989.004.123
Blenniidae	Scartella cristata	MZUSP55440	CIUFSC1574	BLEN 1979.001.120
Bothidae	Bothus ocellatus	MZUSP55378	-	-
Callionymidae	Callionymus bairdi	MZUSP55455	_	-
Carangidae	Caranx crysos	MZUSP55363	-	_
Carangidae	Caranx hippos	-	CIUFSC1311	CARA 1984.012.416
Carangidae	Caranx latus	_	CIUFSC734	CARA 1989.051.455
Carangidae	Chloroscombrus chrysurus	-	CIUFSC1309	CARA 1980.002.406
Carangidae	Oligoplites saliens	_	-	CARA 1988.033.437
Carangidae	Oligoplites saurus	_	CIUFSC727	CARA 2005.072.476
Carangidae	Pseudocaranx dentex	MZUSP55438	-	_
Carangidae	Selene setapinnis	_	CIUFSC379	CARA 2003.073.477
Carangidae	Selene vomer	_	CIUFSC371	-
Carangidae	Trachinotus carolinus	-	CIUFSC723	CARA 2005.077.713
Carangidae	Trachinotus falcatus	_	CIUFSC735	CARA 1988.045.449
Carangidae	Trachinotus marginatus	-	-	CARA 1989.056.460
Carcharhinidae	Carcharhinus brevipinna	_	CIUFSC1357	1 - 1
Carcharhinidae	Carcharhinus isodon	_	CIUFSC1139	_
Carcharhinidae	Carcharhinus obscurus	-	CIUFSC 468	_
Carcharhinidae	Carcharhinus plumbeus	_	CIUFSC1162	_
Carcharhinidae	Carcharhinus porosus	_	CIUFSC481	1
Carcharhinidae	Carcharhinus signatus		CIUFSC1152	_
Carcharhinidae	Rhizoprionodon lalandii	_	CIUFSC1376	-
Centropomidae	Centropomus undecimalis	_	CIUFSC679	CENTRO 1998.018.157
Centropomidae	Centropomus parallelus	-	CIUFSC1513	CENTRO 2002.027.166
Chaenopsidae	Emblemariopsis signifer	MZUSP55448	CIUFSC608	-
Chaetodontidae	Chaetodon striatus	MZUSP55342	CIUFSC1370	_
Chaetodontidae	Prognathodes guyanensis	MZUSP49096	-	_

Continued

Table A1. Continued.

Family	Species	MZUSP	CIUFSC	NEMAR
Clupeidae	Harengula clupeola	<u> </u>	CIUFSC545	CLUP 1980.001.788
Clupeidae	Opisthonema oglinum	-	CIUFSC55	CLUP 2005.042.829
Clupeidae	Sardinella brasiliensis	-	CIUFSC1064	CLUP 1982.004.491
Dactylopteridae	Dactylopterus volitans	MZUSP55431	-	DACTY 1989.003.390
Dactyloscopidae	Dactyloscopus crossotus	MZUSP46668	-	±
Dasyatidae	Dasyatis hypostigma (say)	-	CIUFSC511	-
Diodontidae	Chilomycterus reticulatus	-	CIUFSC259	-
Diodontidae	Chilomycterus spinosus spinosus	MZUSP940	-	DIODS 1999.002.1104
Echeneidae	Echeneis naucrates	-	CIUFSC578	_
Eleotridae	Eleotris pisonis	-	CIUFSC1652	-
Ephippidae	Chaetodipterus faber	MZUSP13314	CIUFSC771	EPHI 1988.006.401
Epinephelidae	Epinephelus marginatus	MZUSP55334	CIUFSC797	-
Epinephelidae	Hyporthodus niveatus	-	CIUFSC1063	-
Epinephelidae	Mycteroperca acutirostris	MZUSP55325	-	_
Epinephelidae	Mycteroperca bonaci	MZUSP55423	_	-
Fistulariidae	Fistularia petimba	-	CIUFSC778	_
Fistulariidae	Fistularia tabacaria	_	CIUFSC530	FISTU 1993.001.787
Gerreidae	Diapterus auratus		CIUFSC115	GERRE 2010.065.1031
Gerreidae	Diapterus rhombeus		CIUFSC62	GERRE 1997.041.1007
Gerreidae Gerreidae			CIUFSC639	GERRE 1997.041.1007 GERRE 1988.024.990
Gerreidae Gerreidae	Eucinostomus argenteus		CIUFSC639 CIUFSC247	GERRE 1988.024.990 GERRE 1987.019.985
	Eucinostomus gula	Ī		
Gerreidae	Eucinostomus melanopterus	-	CIUFSC1816	GERRE 1988.010.976
Gerreidae	Eugerres brasilianus	-	CIUFSC1555	GERRE 1984.008.974
Gobiesocidae	Gobiesox barbatulus (strumosus)	-	CIUFSC1383	GOBIE 1993.001.111
Gobiesocidae	Tomicodonaustralis (fasciatus)	USNM 88042*	CIUFSC536	-
Gobiidae	Barbulifer ceuthoecus	MZUSP55331	-	-
Gobiidae	Bathygobius soporator	-	CIUFSC675	GOBI 1986.019.1051
Gobiidae	Coryphopterus glaucofraenum	MZUSP55332	-	-
Gobiidae	Ctenogobius stigmaticus	-	-	GOBI 1988.036.1068
Gobiidae	Elacatinus figaro	MZUSP49139	-	-
Gobiidae	Microgobius meeki	-	-	GOBI 1982.002.1034
Haemulidae	Anisotremus surinamensis	MZUSP67863	CIUFSC628	HAEMU 1993.017.1156
Haemulidae	Anisotremus virginicus	-	CIUFSC647	-
Haemulidae	Haemulon aurolineatum	MZUSP55357	CIUFSC645	HAEMU 1984.004.1143
Haemulidae	Haemulon bonariense	MZUSP55354	-	-
Haemulidae	Orthopristis ruber	-	CIUFSC396	HAEMU 1988.006.1145
Haemulidae	Pomadasys corvinaeformis	-	CIUFSC400	HAEMU 2010.024.1163
Hemiramphidae	Hemiramphus brasiliensis	-	CIUFSC780	HEMI 1988.004.1170
Hemiramphidae	Hyporhamphus unifasciatus	-	CIUFSC1070	HEMI 1986.001.1167
Holocentridae	Holocentrus adscensionis	MZUSP55390	CIUFSC762	_
Holocentridae	Myripristis jacobus	MZUSP55388	CIUFSC759	-
Kyphosidae	Kyphosus vaigiensis	MZUSP55335	CIUFSC753	-
Labridae	Bodianus pulchellus	MZUSP55398	-	-
Labridae	Halichoeres poeyi	-	CIUFSC823	-
Labridae	Xyrichtys novacula	_	CIUFSC1058	_
Labridae -Scarine	Cryptotomus roseus	MZUSP55425	-	-
Labrisomidae	Labrisomus cricota	MZUSP55339	_	_
Labrisomidae	Labrisomus nuchipinnis	MZUSP55379	CIUFSC599	LABRE 1994.001.1166
Labrisomidae	Malacoctenus aff. triangulatus	MZUSP55450	CIOI 3C399	EABRE 1994.001.1100
	Starksia brasiliensis	MZUSP55367	-	-
Labrisomidae		WIZUSP33307	-	-
Lutjanidae	Lutjanus analis	-	-	LUTJA 1989.002.1178
Lutjanidae	Lutjanus jocu	- M714CDEE 43.0	-	LUTJA 1987.001.1177
Malacanthidae	Malacanthus plumieri	MZUSP55430	-	
Monacanthidae	Monacanthus ciliatus	-	-	MONA 1992.006.1192
Monacanthidae	Stephanolepis hispidus	MZUSP55326	CIUFSC815	MONA 1987.001.1187
Mugilidae	Mugil curema	-	CIUFSC1067	MUGI 1986.001.716
Mugilidae	Mugil liza	-	CIUFSC1967	MUGI 1987.010.725
Mullidae	Pseudupeneus maculatus	MZUSP55344	CIUFSC810	-
Muraenidae	Echidna catenata	MZUSP16623	-	-
Muraenidae	Gymnothorax moringa	-	CIUFSC556	<u>-</u>
Muraenidae	Gymnothorax ocellatus	-	CIUFSC541	MURAE 1984.001.1199

Continued

 Table A1. Continued.

Family	Species	MZUSP	CIUFSC	NEMAR
Muraenidae	Gymnothorax vicinus	4	CIUFSC554	-
Narcinidae	Narcine brasiliensis	-	CIUFSC576	NARCI 1991.001.1279
Ogcocephalidae	Ogcocephalus vespertilio	MZUSP55433	CIUFSC575	OGCOC 1990.001.1276
Ophichthidae	Myrichthys breviceps	MZUSP55337	-	-
Ophichthidae	Myrichthys ocellatus	-	CIUFSC557	-
Ophidiidae	Ophidion holbrooki	-	CIUFSC1524	-
Ostraciidae	Acanthostracion quadricornis	MZUSP49101	CIUFSC531	-
Ostraciidae	Lactophrys trigonus	MZUSP55428	-	OSTRA 2001.001.1124
Paralichthyidae	Paralichthys brasiliensis	-	CIUFSC437	-
Paralichthyidae	Syacium micrurum	-	CIUFSC806	-
Pempheridae	Pempheris schomburgki	-	CIUFSC1060	-
Polynemidae	Polydactylus virginicus	-	-	POLY 2011.001.1218
Pomacanthidae	Holacanthus ciliaris	MZUSP55392	CIUFSC1061	-
Pomacanthidae	Pomacanthus paru	MZUSP55458	CIUFSC822	-
Pomacentridae	Abudefduf saxatilis	-	CIUFSC1368	POMAC 1991.001.1112
Pomacentridae	Chromis jubauna	MZUSP55432	-	-
Pomacentridae	Chromis multilineata	-	CIUFSC1059	-
Pomacentridae	Stegastes fuscus	MZUSP55343	CIUFSC620	-
Pomacentridae	Stegastes pictus	MZUSP55346	-	_
Pomacentridae	Stegastes variabilis	MZUSP55400	CIUFSC610	_
Pomatomidae	Pomatomus saltatrix	-	CIUFSC1545	POMA 1991.018.1101
Priacanthidae	Cookeolus japonicus	-	-	PRIAC 2005.002.1174
Priacanthidae	Priacanthus arenatus	_	CIUFSC769	PRIAC 2011.004.1176
Rachycentridae	Rachycentron canadum	_	CIUFSC1491	-
Rhinobatidae	Rhinobatos percellens	_	-	RHINO 2005.002.1279
Rhinobatidae	Zapteryx brevirostris	_	CIUFSC503	RHINO 2005.003.1280
Sciaenidae	Menticirrhus americanus		CIUFSC1965	SCIA 1999,110.612
Sciaenidae	Menticirrhus dinericanus Menticirrhus littoralis		CIUFSC683	SCIA 1991.100.592
Sciaenidae	Micropogonias furnieri		CIUFSC1831	SCIA 1988.084.576
Sciaenidae	Odontoscion dentex	MZUSP55345	CIUFSC710	3CIA 1900.004.370
Sciaenidae				-
	Pareques acuminatus Scomberomorus brasiliensis	MZUSP55386	CIUFSC601	- CCOMP 2011 001 1104
Scombridae		•	-	SCOMB 2011.001.1194
Scorpaenidae	Scorpaena isthmensis		- CUJECC1272	SCORP 1982.001.1136
Scorpaenidae	Scorpaena plumieri	-	CIUFSC1373	SCORP 1987.002.1137
Serranidae	Diplectrum formosum	-	-	SERRA 1986.003.1285
Serranidae	Diplectrum radiale	- NATUSDAGGAO(A)	-	SERRA 1982.001.1283
Serranidae	Rypticus randalli	MZUSP46648(1)	-	-
Serranidae	Serranus flaviventris	MZUSP55429	-	_
Sparidae	Archosargus rhomboidalis	-	CIUFSC1072	SPAR 1989.012.774
Sparidae	Calamus pennatula	MZUSP69960	-	
Sparidae	Diplodus argenteus	MZUSP66545	CIUFSC1366	SPAR 1988.006.768
Sparidae	Pagrus pagrus	MZUSP70093	CIUFSC624	-
Sphyraenidae	Sphyraena guachancho	-	CIUFSC1840	SPHYR 1983.001.165
Sphyraenidae	Sphyraena tome	-	UFRGS 04429	-
Sphyrnidae	Sphyrna lewini	-	CIUFSC1134	-
Sphyrnidae	Sphyrna zygaena	-	CIUFSC1155	-
Syngnathidae	Hippocampus patagonicus	-	CIUFSC315	-
Syngnathidae	Hippocampus reidi	MZUSP55456	-	SYNG 1981.001.488
Syngnathidae	Microphis lineatus	-	CIUFSC275	-
Syngnathidae	Sgnathus folleti	-	-	SYNG 1989.002.489
Synodontidae	Synodus bondi (foetens)	-	-	SYNO 1983.001.478
Tetraodontidae	Canthigaster figueiredoi	MZUSP55355	-	-
Tetraodontidae	Lagocephalus laevigatus	-	CIUFSC1365	TETRA 1991.019.1238
Tetraodontidae	Sphoeroides greeleyi	-	CIUFSC245	TETRA 1980.001.1220
Tetraodontidae	Sphoeroides spengleri	MZUSP55338	CIUFSC552	TETRA 2010.031.1250
Tetraodontidae	Sphoeroides testudineus	_	CIUFSC388	TETRA 1980.004.1223
Tetraodontidae	Sphoeroides tyleri	_	_	TETRA 2004.028.1247
Triglidae	Prionotus nudigula	_	-	TRIGL 2010.024.1275
Triglidae	Prionotus punctatus		CIUFSC1966	TRGL 1989.017.1268

^{*} Tomicodon australis holotype is from São Francisco do Sul, SC, Brazil.